

National Wastewater Surveillance Programme - COVID-19

Weeks 9 & 10 (Weeks ending 5 March & 12 March 2023)

Report prepared on 15 March 2023

100%

sites tested had SARS-CoV-2 detected (83/83 sites) in week 10

66%

NZ population covered by wastewater testing in week 10

**Omicron
XBB (~43%)**

Most prevalent variant detected in week 10

Nationally, SARS-CoV-2 levels have moderately increased but are lower than 12 weeks ago. Variant analysis suggests that XBB and CH.1.1 continue to be the most common variants detected.

- Comparing this week ending 12 March to week ending 5 March 2023, 38% of sites show an increase in SARS-CoV-2 levels while 38% sites showed a decrease in SARS-CoV-2.
- Comparing this week ending 12 March 2023 to one month ago (week ending 12 February 2023), 54% of sites show an increase in SARS-CoV-2 levels while 28% of sites showed a decrease in SARS-CoV-2 levels.
- The main variants detected in wastewater in the week ending 12 March 2023 (week 10) were XBB (includes XBB.1.5, ~43%), CH.1.1 (~28%), BA.2.75* (includes XBF, ~25%). Minor contributions from BQ.1.1 (~2%), and XBC (~2%). BA.4/BA.5 not detected.
- Weather-related impacts in late January and February 2023 resulted in fewer samples being collected in some regions. For example, Northland and Auckland sites were impacted by flooding in late January, and Hawke's Bay and Gisborne by Cyclone Gabrielle in early February. While many sites had recommenced regular sampling by week 10, others continue to be missing (for example South Western Interceptor, Napier and Gisborne).

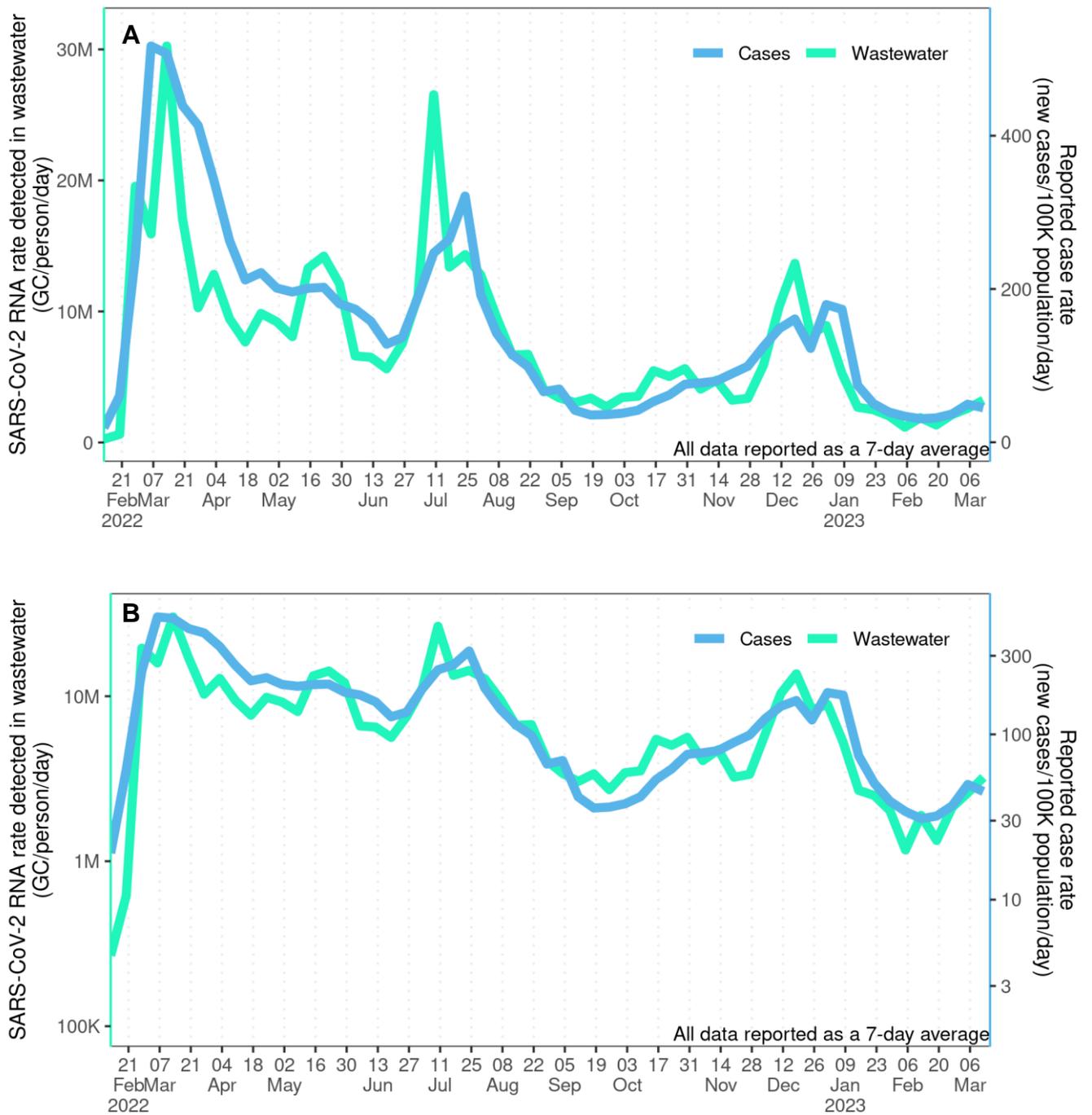


Figure 1. National timeseries of estimated SARS-CoV-2 genome copies (GC) in wastewater rate (GC/person/day, green line) and reported case rate (new cases/100,000 population/day, blue line) on a linear scale (A) and Log₁₀ scale (B). Data reported as 7-day average.

Results for Weeks 9 & 10 (Weeks ending 5 March & 12 March 2023)

In the two weeks ending 12 March 2023, 221 samples were collected from 83 locations across New Zealand.

SARS CoV-2 RNA was detected in 217/221 (98%) samples from 83/83 (100%) sites (Figure 2, Table 1).

No sampling from the Auckland South Western Interceptor has been possible from week 5 following subsidence to the surrounding ground around the sampler during the Auckland flooding. Currently, there is no safe access point to sample from the South Western Interceptor. A combined sample from Mangere treatment plant is being collected that includes the South Western catchment.

In week 10, sampling from all seven regular Northland sites had recommenced (4 sites collected in week 9). No samples from Gisborne and Napier were collected in weeks 9 and 10 (nor in week 8).

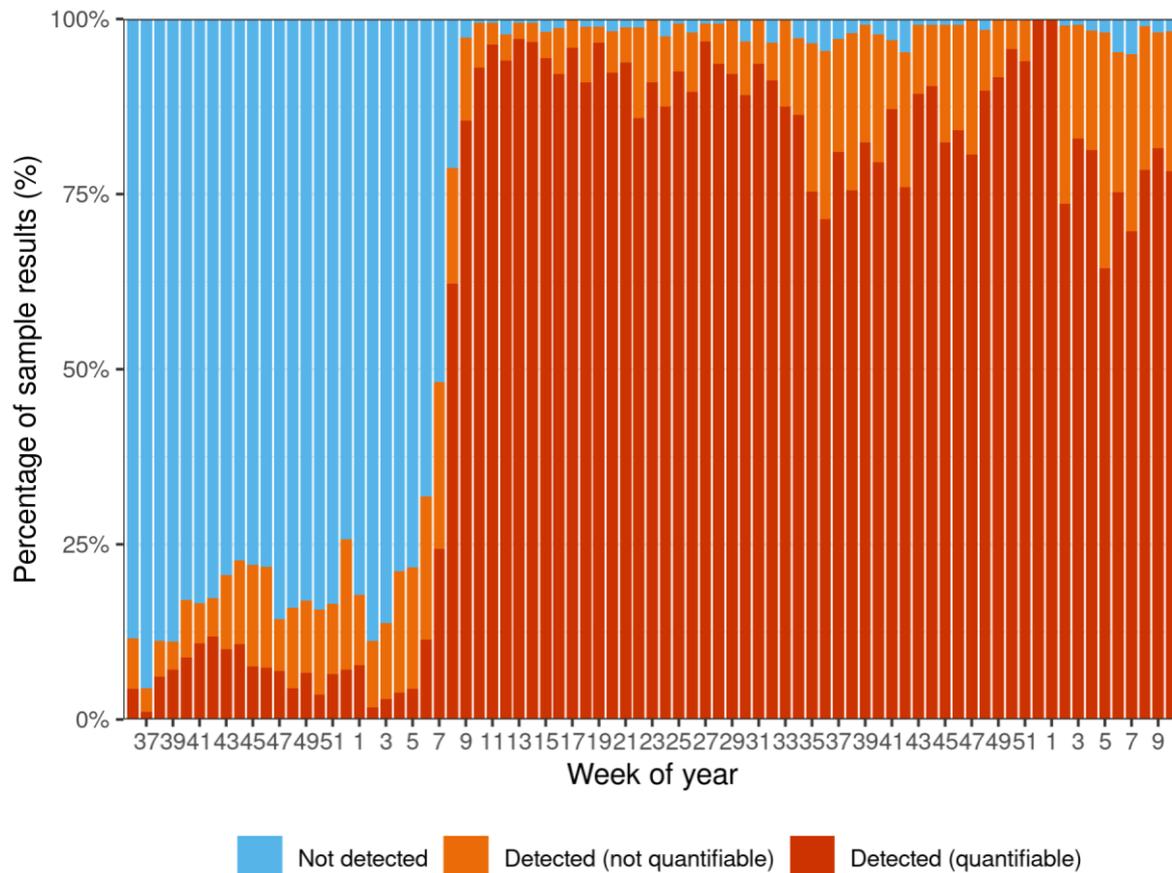


Figure 2. Results for SARS-CoV-2 RNA in wastewater collected across New Zealand.

Regional Trends

Regional summaries (Figure 3) of the wastewater data indicates generally steady or increasing viral levels in all regions in week 10 compared to previous weeks. Note that regional trend analysis for week 52 (2022) and week 1 (2023) was only possible for Auckland Metro, as there were limited samples collected during the holiday period. Viral quantitation for the other regions were therefore not available during this period (denoted by dashed line).

Due to the weather-related impacts in February 2023, fewer samples were collected in some regions. The Central regional summary excludes Hawke’s Bay samples in weeks 8-10, and analysis for the Northern region was not possible in between weeks 7-9 due to too few samples being received from this region. The dashed blue line shows inferred level in this region during this time (Figure 3).

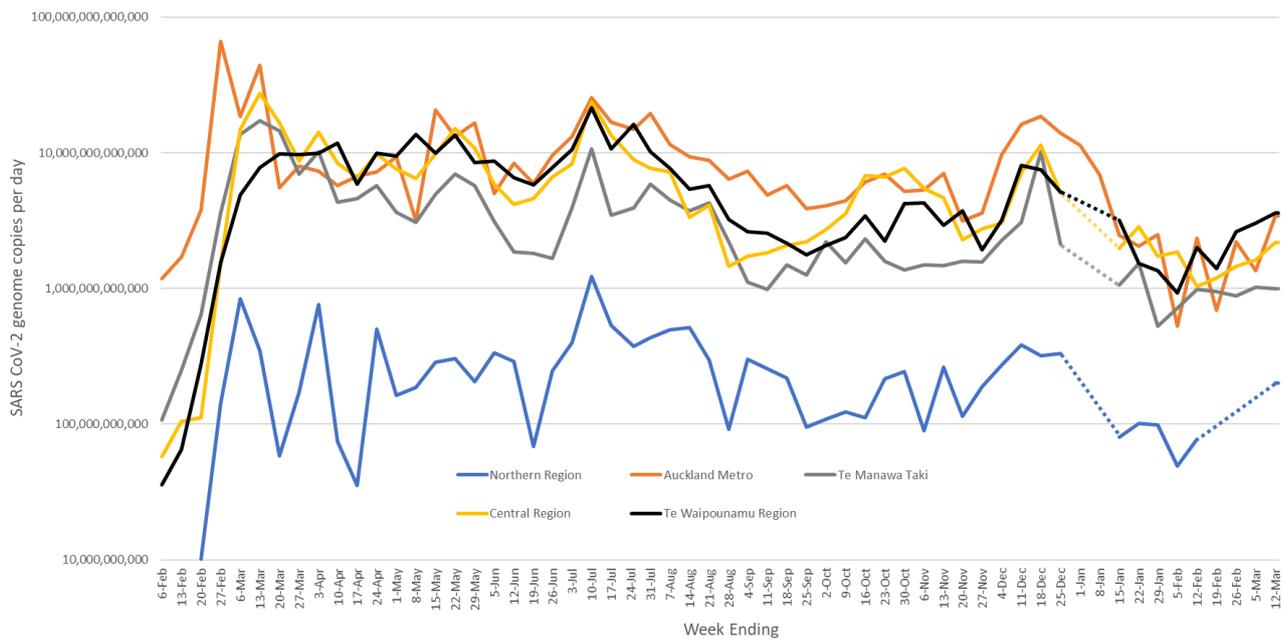


Figure 3. Total SARS-CoV-2 genome copies detected per day in the five Ministry of Health regions. Dashed lines are inferred levels during periods when samples were either not collected (Christmas period) or insufficient numbers collected (due to weather impacts) for the region.

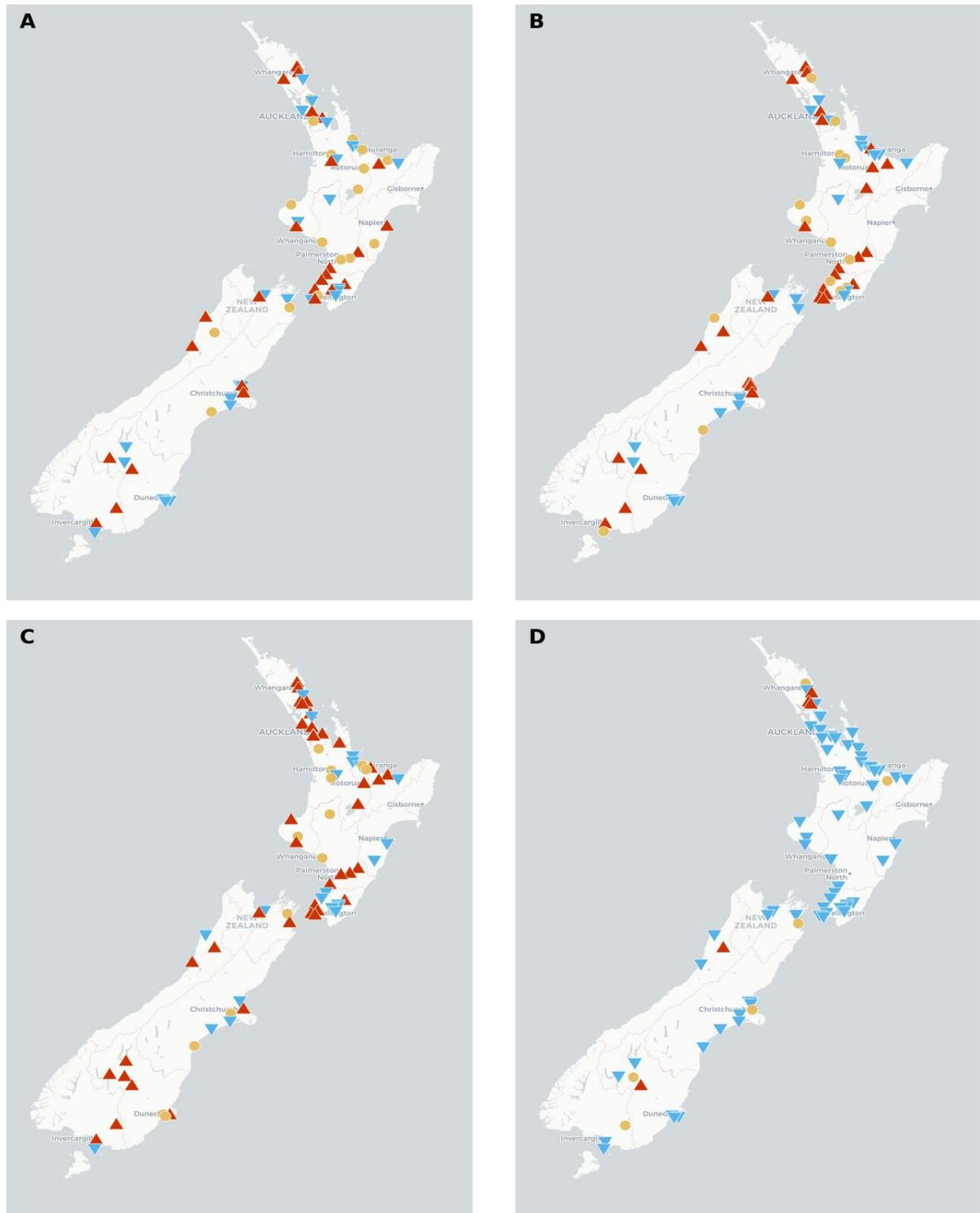


Figure 4. Comparison of SARS-CoV-2 levels for the week ending 12 March 2023, compared to levels measured: A) 1 week ago; B) 2 weeks ago; C) 4 weeks ago; D) 12 weeks ago. Only sites with results for both time points are included. When the viral quantity is 30% or more higher this is labelled as increased (red up arrow on map). When the viral quantity is 30% or more lower, this is labelled as decreased (blue down arrow on map). If viral levels have changed less than this in the compared weeks, this is labelled as no change (yellow circle on map). Interactive map of weekly results available publicly at <https://www.poops.nz/>

Wastewater Variant Analysis

In collaboration with Wilderlab, ESR generated the variant analysis results (Table 1, Figure 5) from sentinel sites in week 9 (ending 5 March 2023) and week 10 (ending 12 March 2023).

Wastewater variant analysis is based on sequencing a short fragment of the spike gene and therefore provides less resolution than WGS from clinical cases. As such, some specific lineages cannot be distinguished from each other, and are reported as variant groups. The following variants/groups are reported: BA.4/BA.5, BA.2.75* (includes BA.2.75/XBF/BR.2 subvariants), CH.1.1, BQ.1.1, XBB (includes XBB.1.5) and XBC.

Due to the increasing complexity of variants in the population, each at relatively low levels, the current approach for sequencing wastewater samples needs to be more precise to report percentages for each variant at the sentinel site level. Instead, the presence of each lineage will currently be reported. ESR is actively testing and developing methods to address the current uncertainty and increase the resolution to identify variants in wastewater.

Consistent with the WGS of clinical cases, the CH.1.1 subvariant will now be reported separately from other BA.2.75* subvariants.

Results for weeks 9 and 10

CH.1.1 was **frequently detected** in weeks 9 (14/17 sites) and 10 (14/18 sites). CH.1.1 comprised ~41% of sequencing reads nationally in week 9, declining to ~28% sequencing reads in week 10. Other subvariants in the BA.2.75* group (including BM.4, BR.2, XBF and BA.2.75) accounted for another ~4% reads in week 9. This rose to ~25% of sequencing reads nationally in week 10.

XBB (includes XBB.1.5) was also **frequently detected**, comprising ~46% of reads nationally in week 9 (14/17 sites), and ~43% of reads nationally in week 10 (15/18 sites).

BQ.1.1 was only identified in ~4% of national sequence reads in week 9 and ~2% in week 10, and with no consistency between weeks in which sites had BQ.1.1 detections (2/17 sites in week 9, 3/18 sites in week 10, Table 1).

The XBC variant also remains at low levels, comprising ~4% (week 9) and ~2% (week 10) of sequencing reads nationally.

The BA.4/BA.5 variant group (including BF.7) was only detected in Palmerston North in week 9 (accounting for ~1% of sequences nationally) and not detected at any site in week 10.

	Week 9						Week 10					
	BA.4/BA.5	BA.2.75*	CH.1.1	BQ.1.1	XBB	XBC	BA.4/BA.5	BA.2.75*	CH.1.1	BQ.1.1	XBB	XBC
Whangarei												
North Shore												
Auckland East												
Auckland Southwest												
Auckland West												
Mt Maunganui												
Tauranga												
Rotorua												
Taupo												
Gisborne												
New Plymouth												
Palmerston North												
Porirua												
Hutt Valley												
Wellington (Moa Point)												
Nelson												
Christchurch												
Queenstown												
Dunedin (Tahuna)												
Dunedin (Mosgiel)												
All Sites (national)	1	4	41	4	46	4	25	28	2	43	2	

Table 1. Data from 17 wastewater sentinel sites sampled in week 9 (ending 5 March 2023) and 18 sentinel sites sampled in week 10 (ending 12 March 2023) using a S-gene (spike) barcoding assay able to ‘call’ the BA.4/BA.5, the BA2.75* constellation (includes BA.2.75/XBF/BR.2 subvariants), CH.1.1, BQ.1.1, XBB (includes XBB.1.5) and XBC (sub)variants. Coloured box denotes that the variant was detected at that site that week, white box denotes that the variant was not detected, and grey box denotes site was not sampled that week. Numbers in the bottom row denote the estimated percentage of each variant at the national scale.

Variant Timeline - National

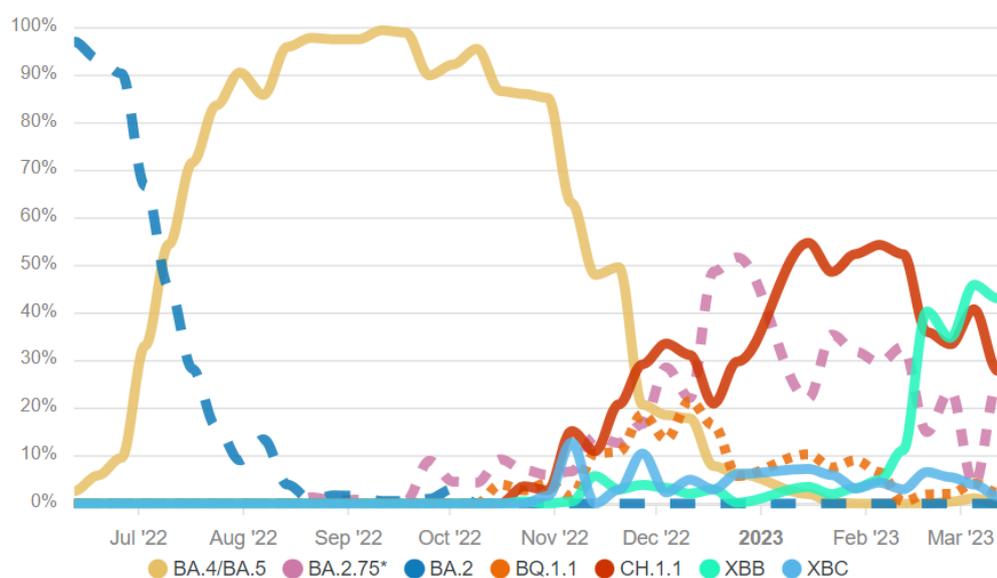
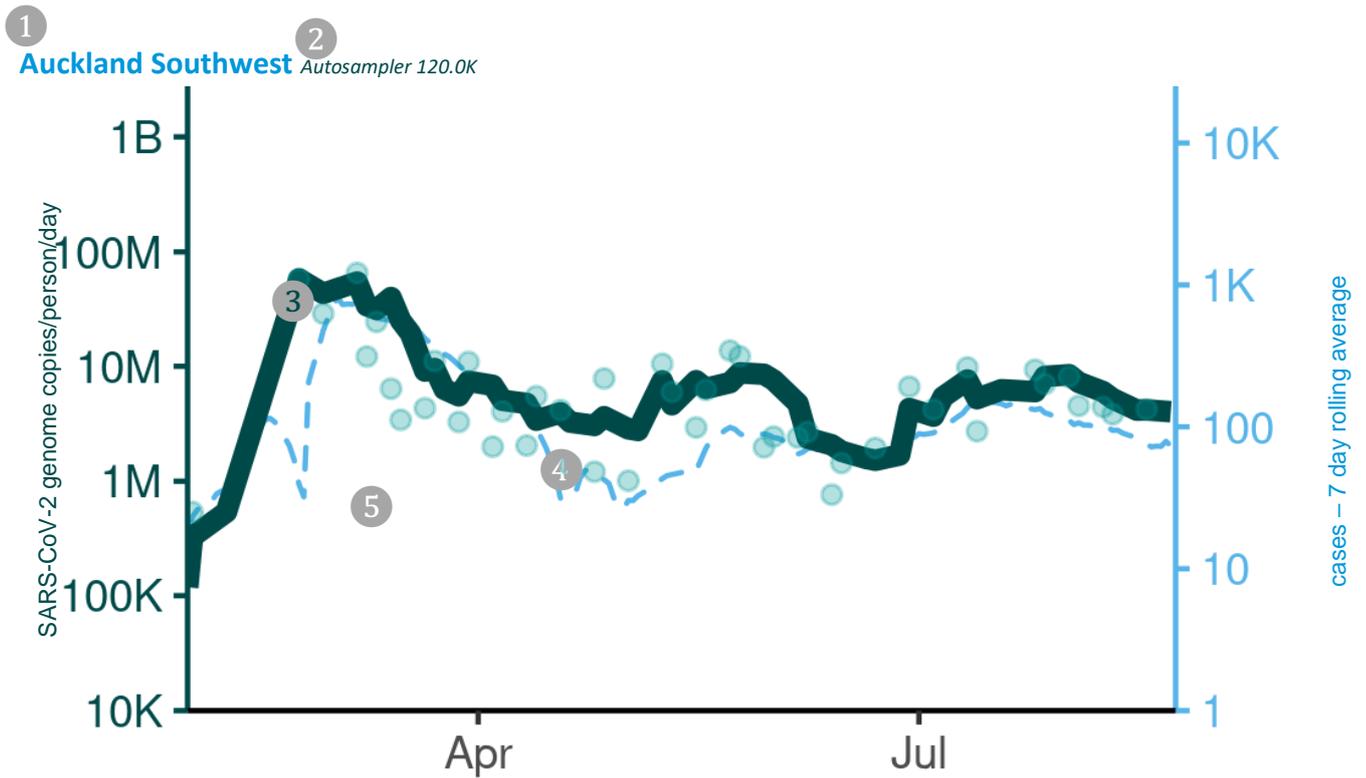


Figure 5. Change in variant prevalence over time at a national scale. Data are collected from up to 20 sentinel sites each week.

Interpreting Sites Graphs

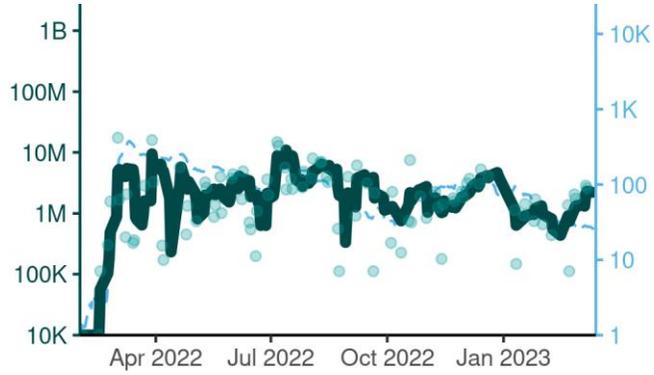


- 1** Site Name
- 2** Sample collection method and population. Results based on autosampler may be more representative than grab sample-based results.
- 3** Wastewater results shown as solid line | 14-day average of genome copies/person/day on a \log_{10} scale.
- 4** Individual results samples shown as circles | Rolling 14-day average of genome copies/person/day on a \log_{10} scale.
- 5** Rolling 7-day average of new cases shown as dashed line | New cases reported in a catchment based on reported date of illness on a \log_{10} scale. This data is not available for all sites and subject to change.

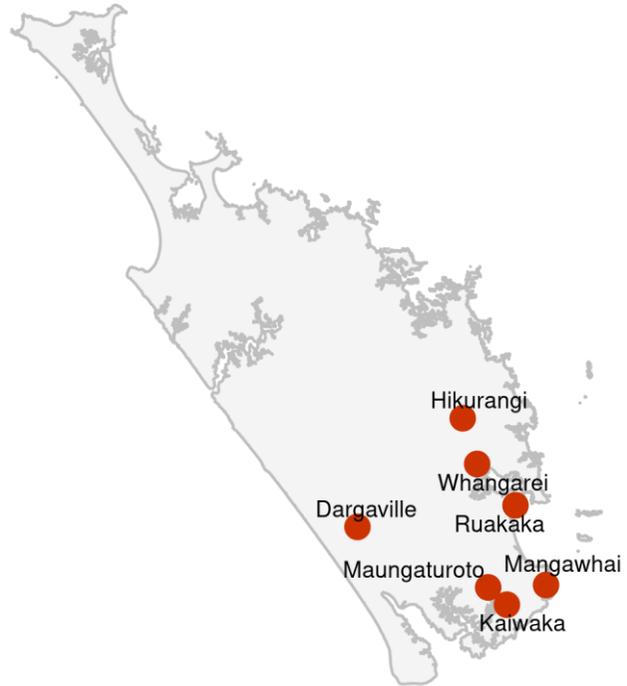
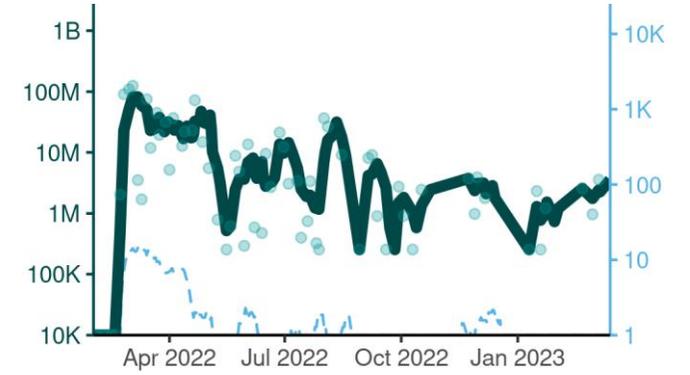
Note: Wastewater and cases data are on a \log_{10} scale. Scales on all graphs have been normalized to cover the same scale on every graph. Care should be taken when interpreting the data.

Northland

Whangarei Autosampler 65.0K

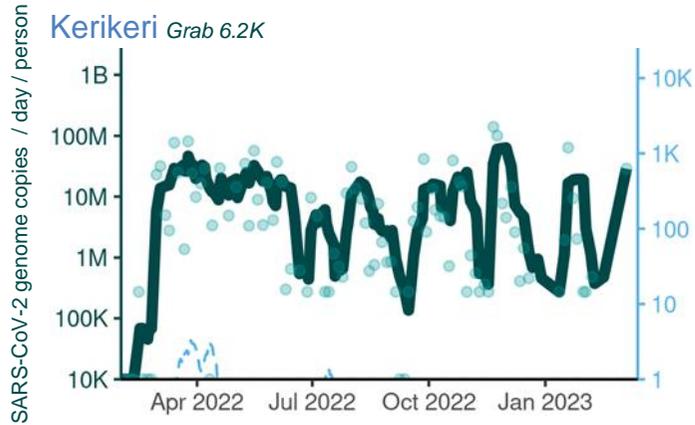


Dargaville Grab 5.0K

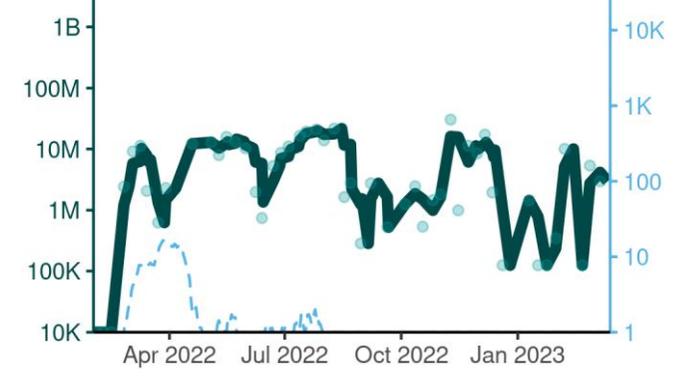


Status ● Detected ● Not detected

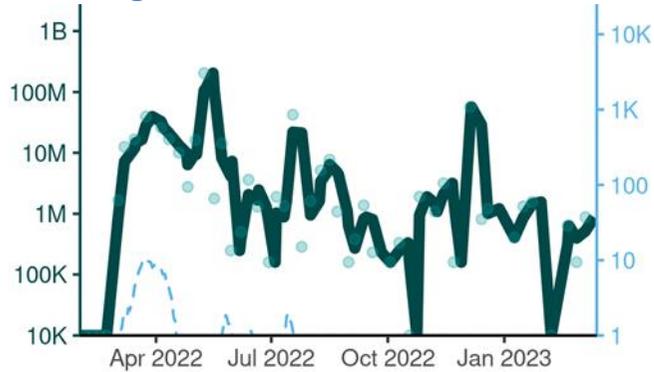
Kerikeri Grab 6.2K



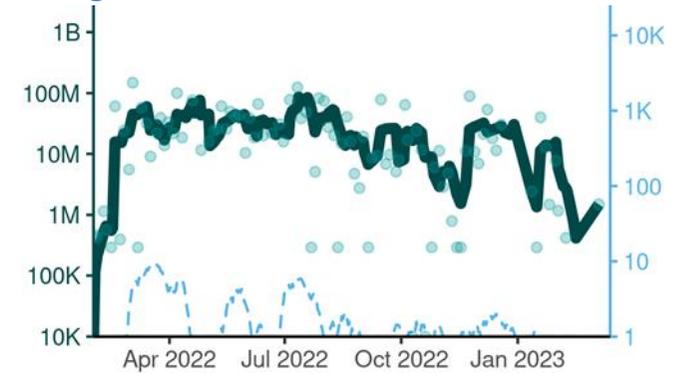
Ruakaka Grab 4.5K



Hikurangi Grab 1.7K

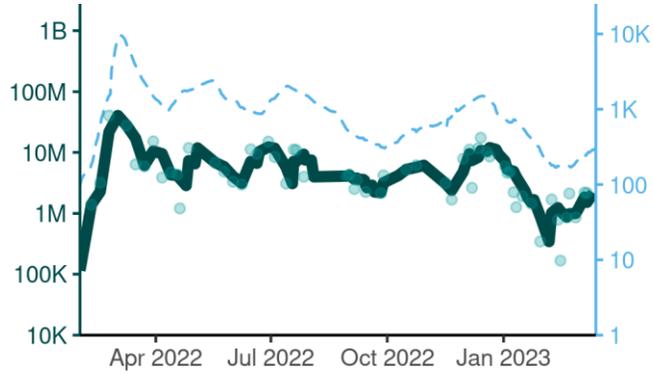


Mangawhai Grab 1.1K

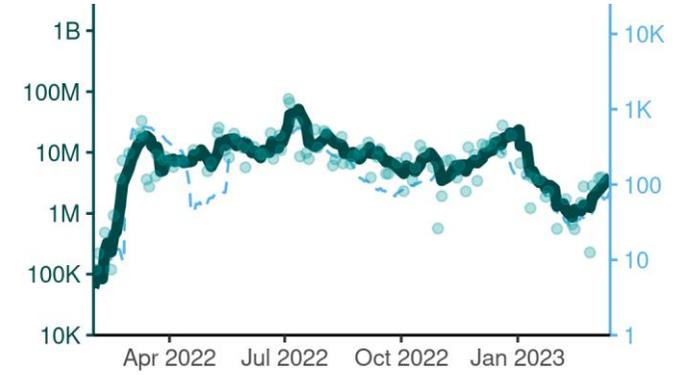


Auckland

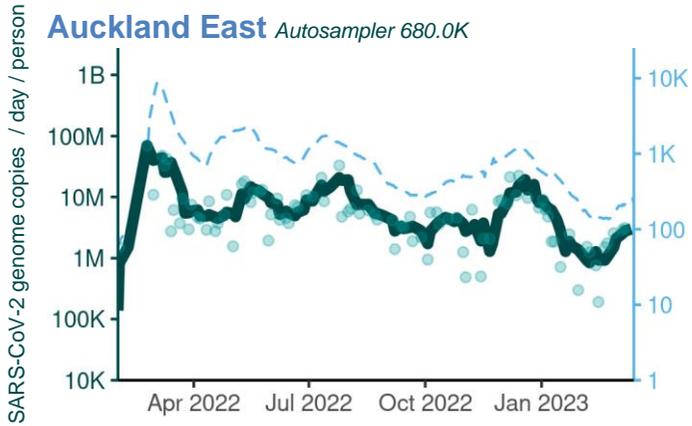
Auckland (Combined) Autosampler 1.1M



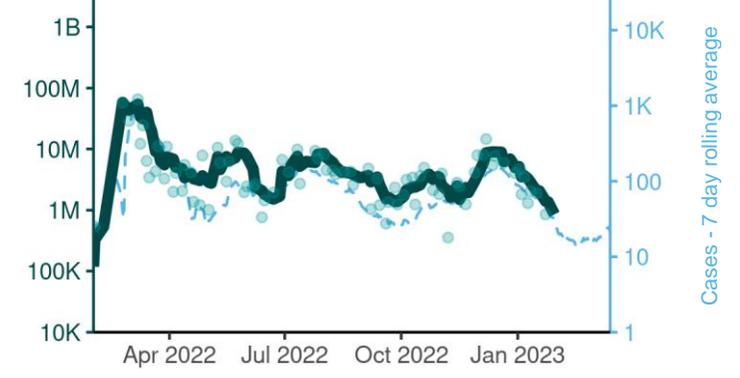
North Shore Autosampler 240.0K



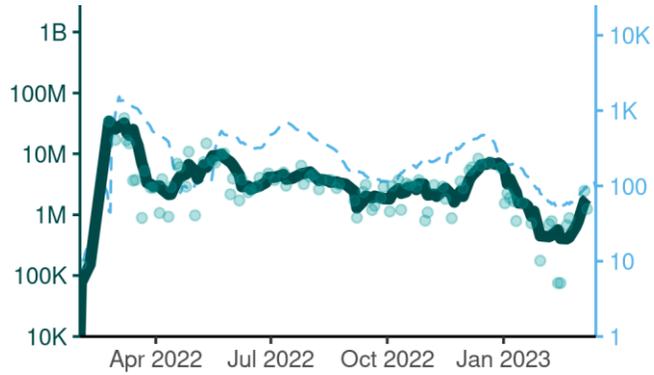
Auckland East Autosampler 680.0K



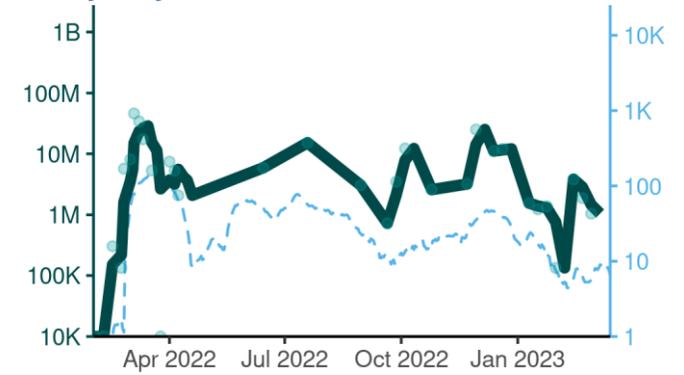
Auckland Southwest Autosampler 120.0K



Auckland West Autosampler 315.0K

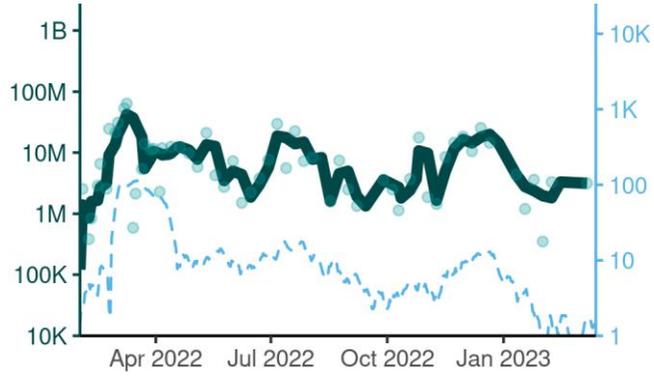


Army Bay Autosampler 42.0K

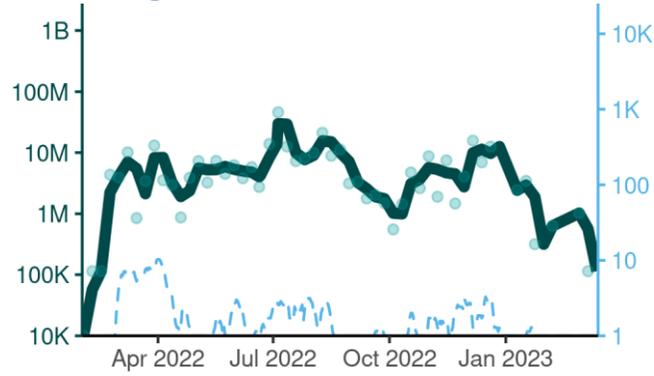


Status ● Detected ● Not detected

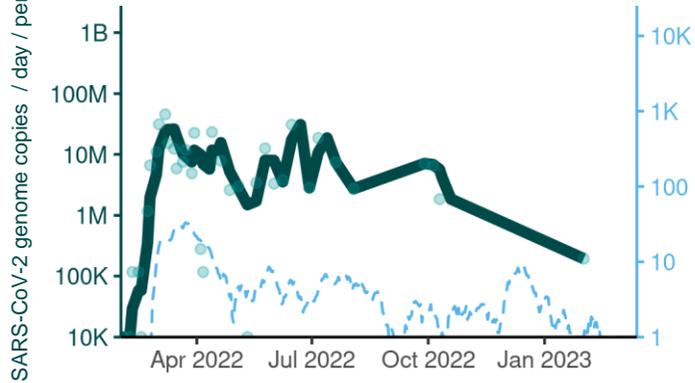
Pukekohe Autosampler 20.9K



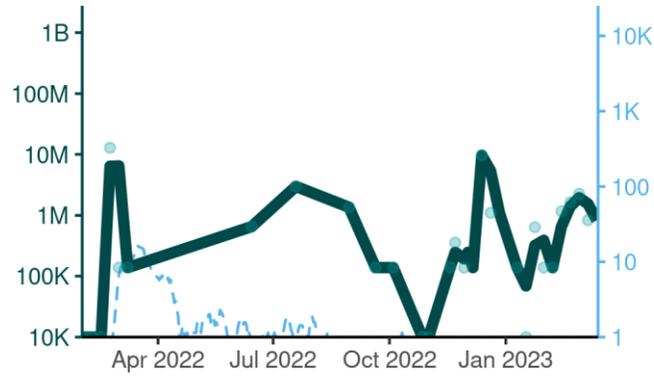
Snells/Algies Autosampler 4.0K



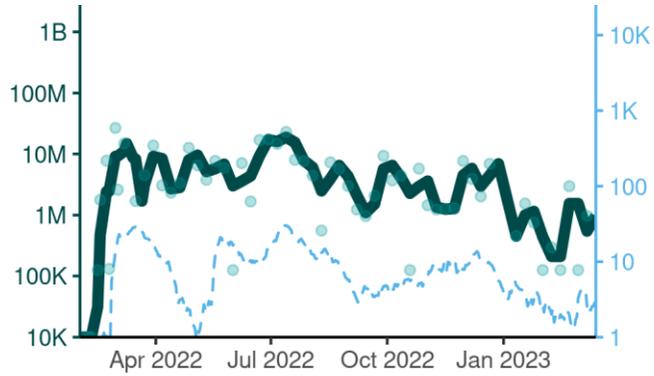
Waiuku Grab 7.9K



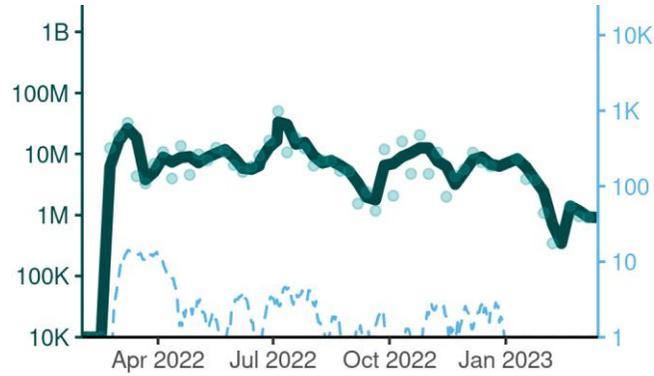
Helensville Autosampler 3.8K



Beachlands Grab 6.8K



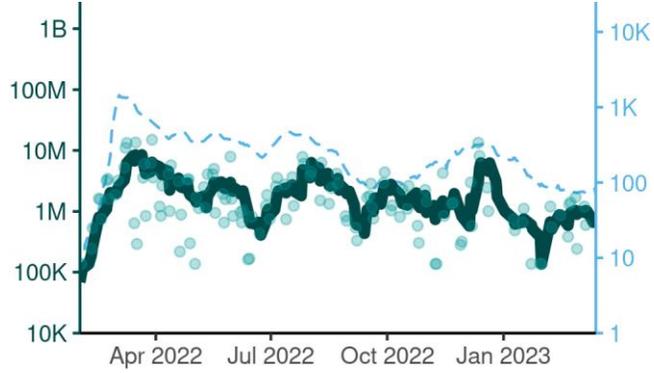
Warkworth Autosampler 3.5K



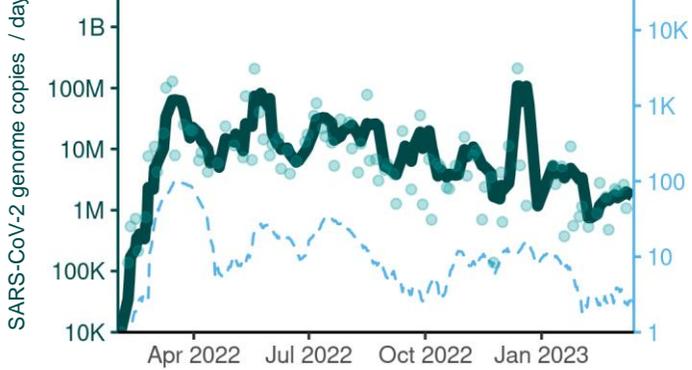
Cases - 7 day rolling average

Waikato

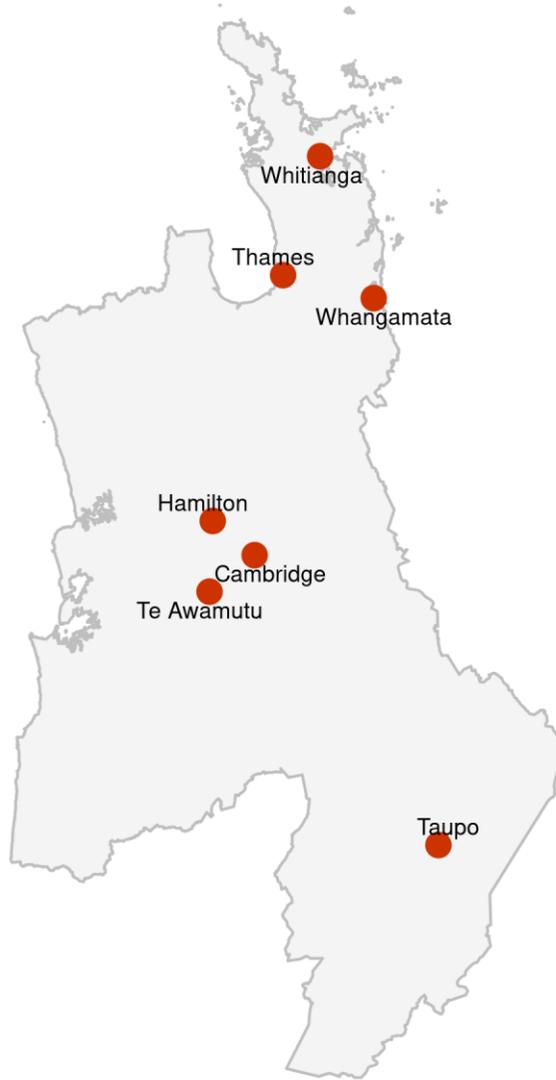
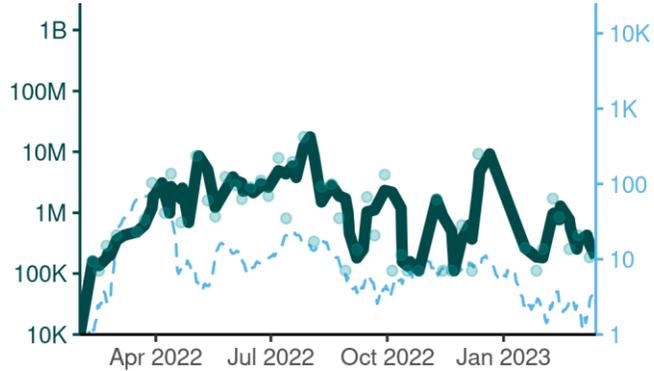
Hamilton Autosampler 169.0K



Taupo Auto/grab 23.0K

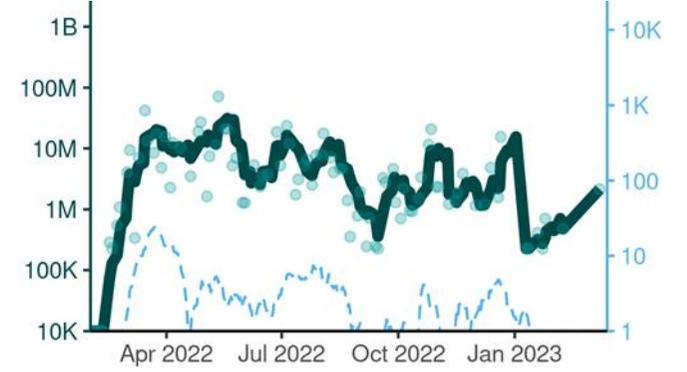


Cambridge Autosampler 20.1K

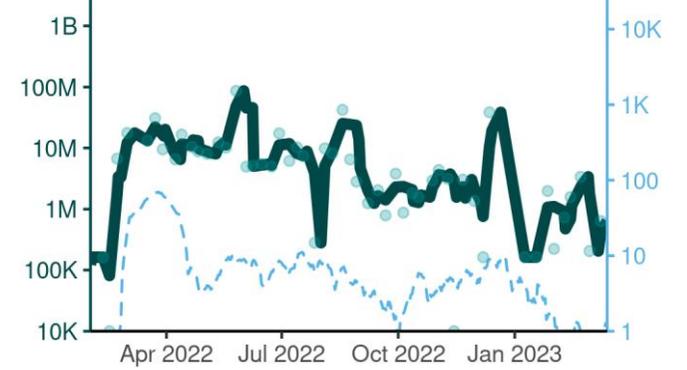


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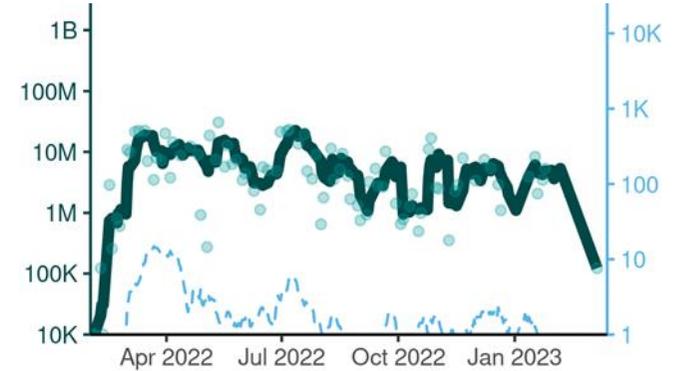
Thames Autosampler 7.5K



Te Awamutu Autosampler 13.1K



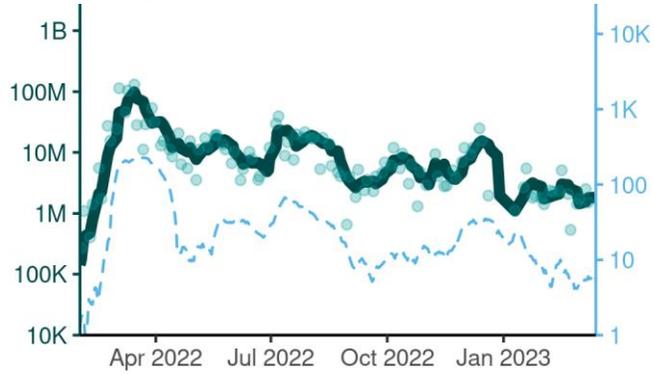
Whitianga Autosampler 6.6K



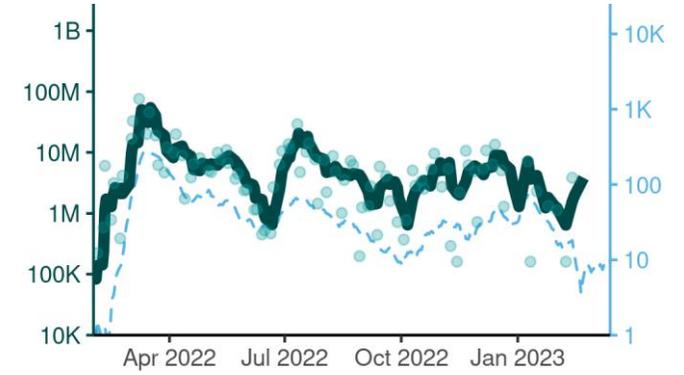
Cases - 7 day rolling average

Bay of Plenty and Gisborne

Mt Maunganui/Papamoa Autosampler 65.0K

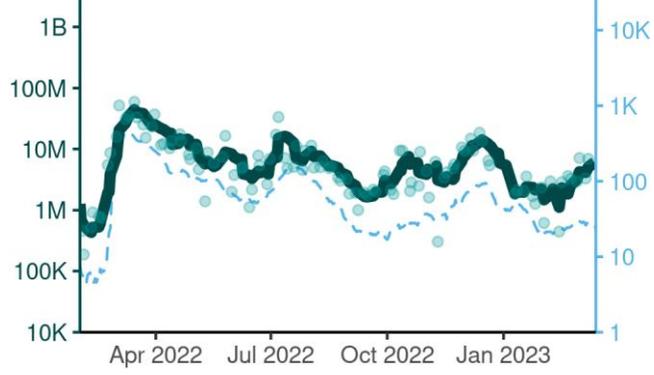


Gisborne Autosampler 37.0K

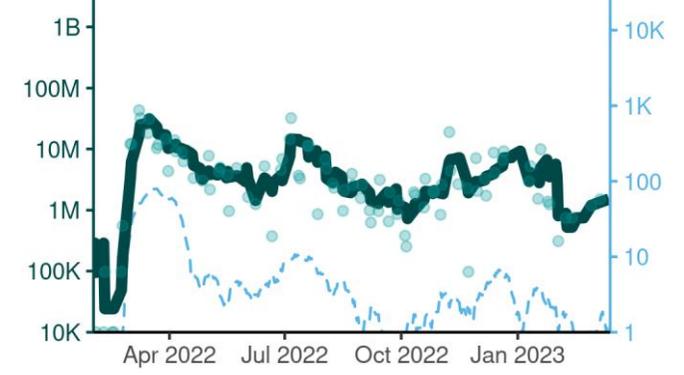


SARS-CoV-2 genome copies / day / person

Rotorua Autosampler 59.0K

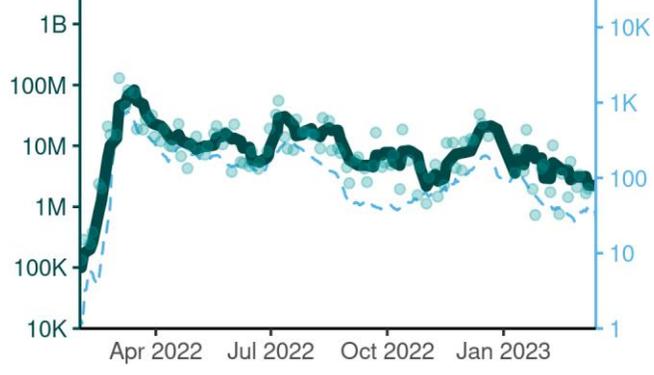


Whakatane Autosampler 21.0K



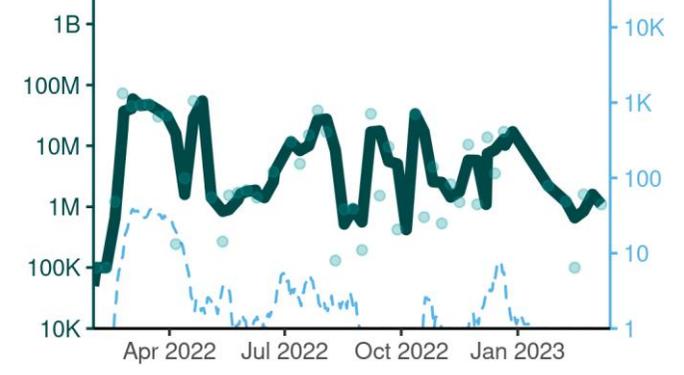
Cases - 7 day rolling average

Tauranga Autosampler 50.0K

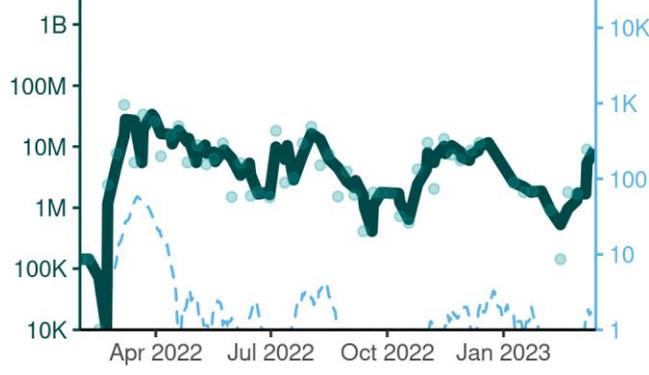


Status ● Detected ● Not detected

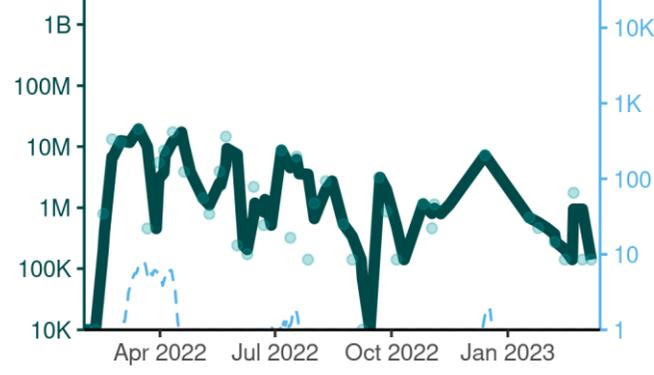
Te Puke Autosampler 9.7K



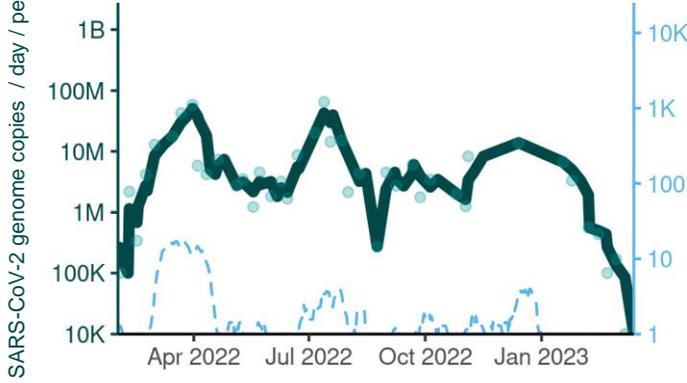
Kawerau Autosampler 7.0K



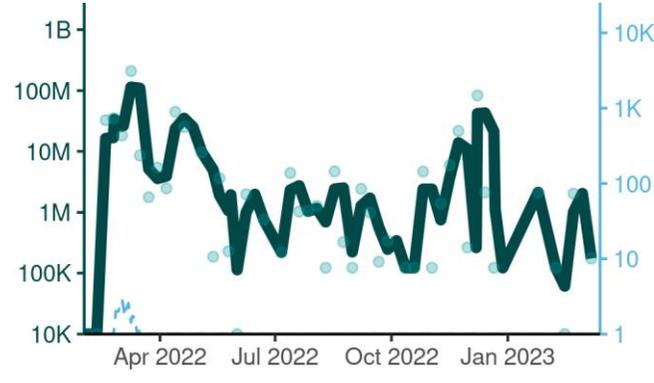
Waihi Beach Autosampler 3.6K



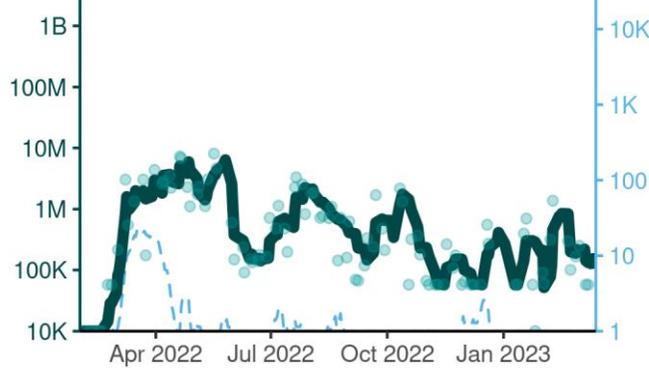
Katikati Autosampler 5.5K



Maketu Autosampler 1.3K



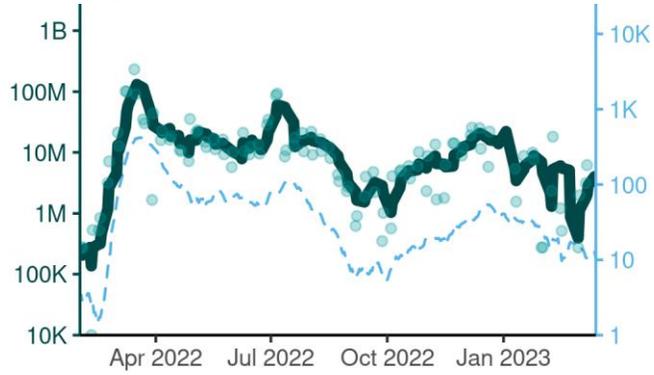
Opotiki Autosampler 3.8K



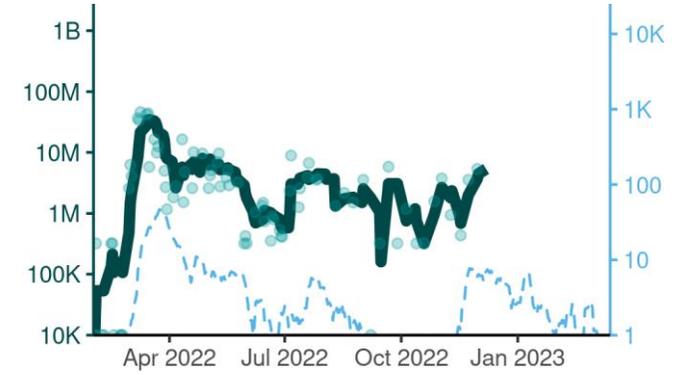
Cases - 7 day rolling average

Hawke's Bay

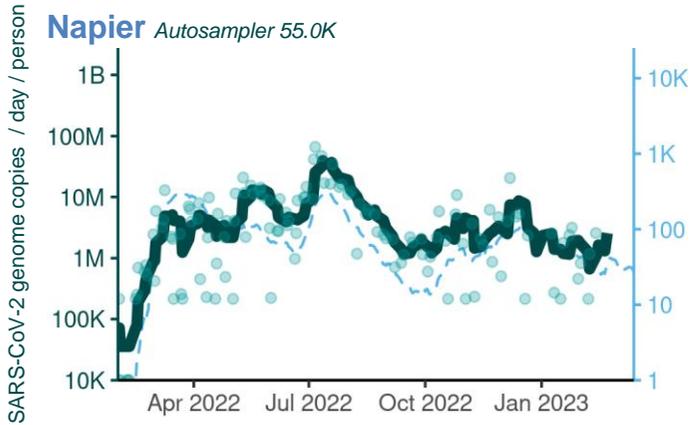
Hastings Autosampler 80.0K



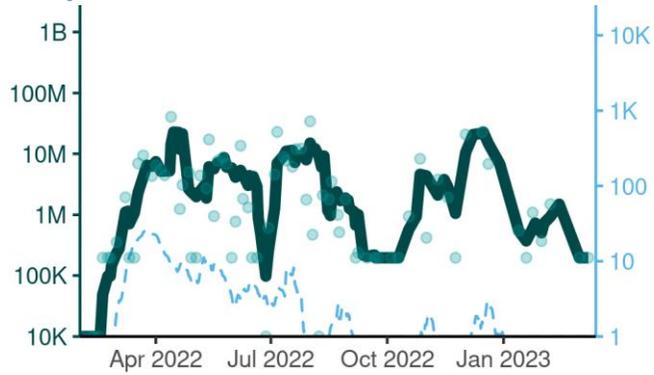
Wairoa Grab 4.4K



Napier Autosampler 55.0K



Waipukurau Autosampler 4.6K

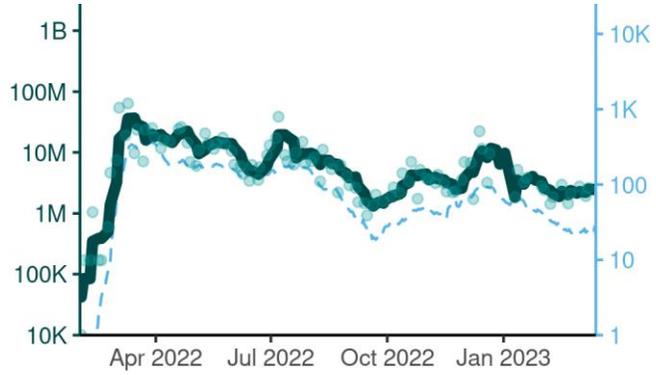


Status ● Detected ● Not detected

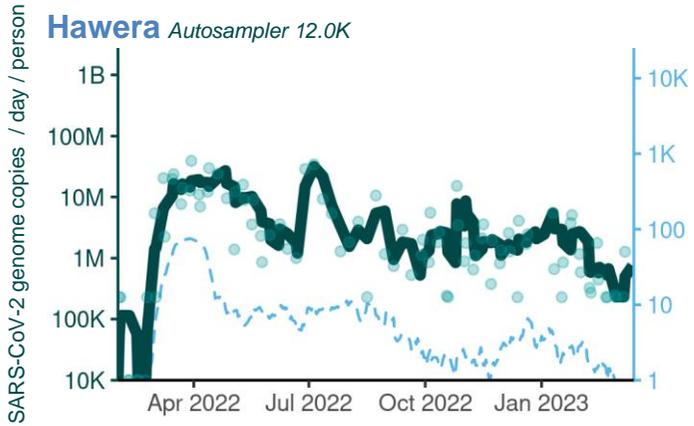
Cases - 7 day rolling average

Taranaki

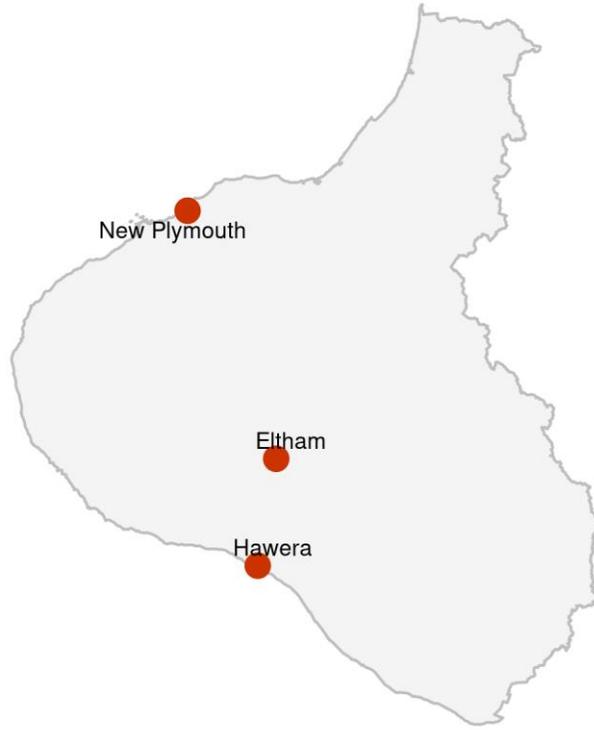
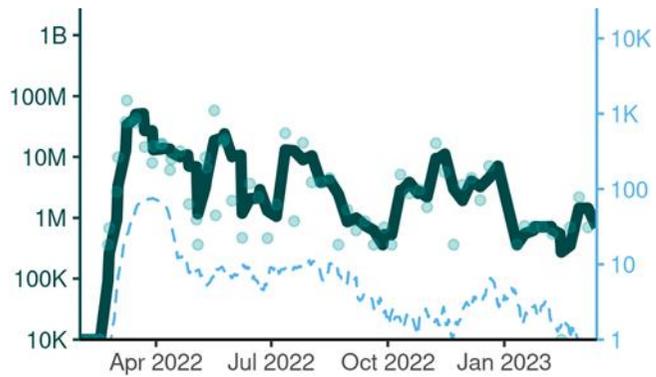
New Plymouth Autosampler 88.0K



Hawera Autosampler 12.0K



Eltham Autosampler 2.0K

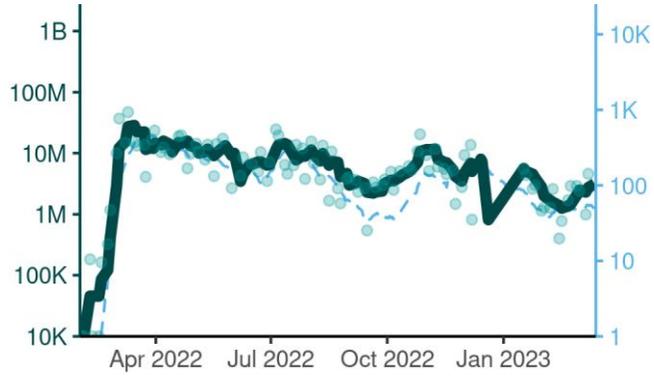


Status ● Detected ● Not detected

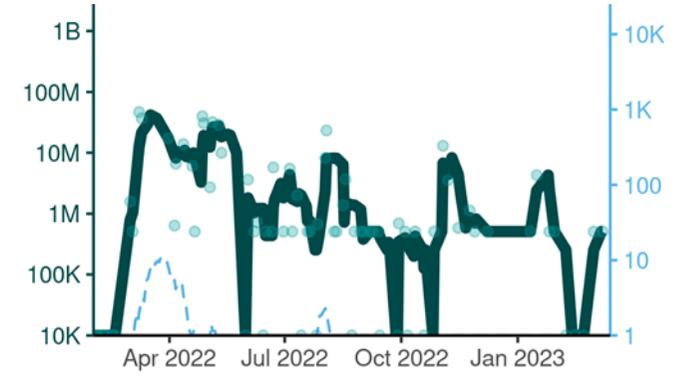
Cases - 7 day rolling average

Manawatu-Whanganui

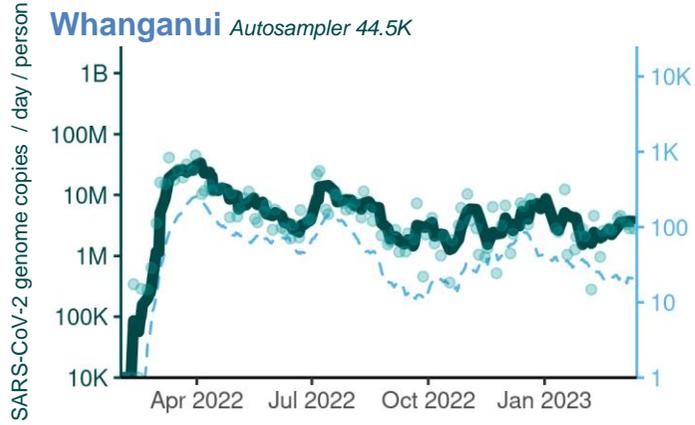
Palmerston North Autosampler 90.0K



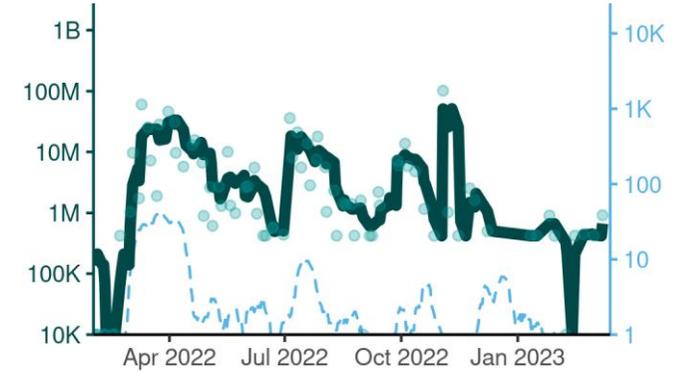
Woodville Grab 1.7K



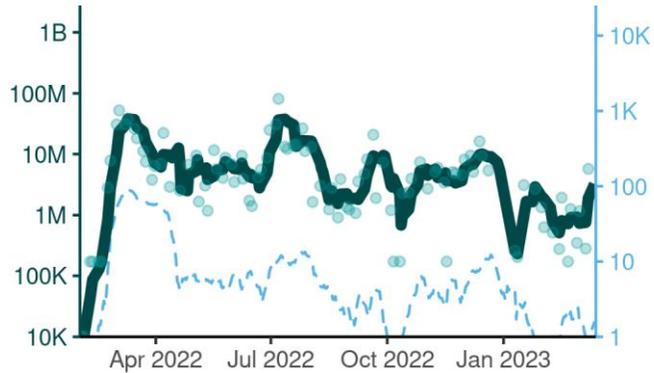
Whanganui Autosampler 44.5K



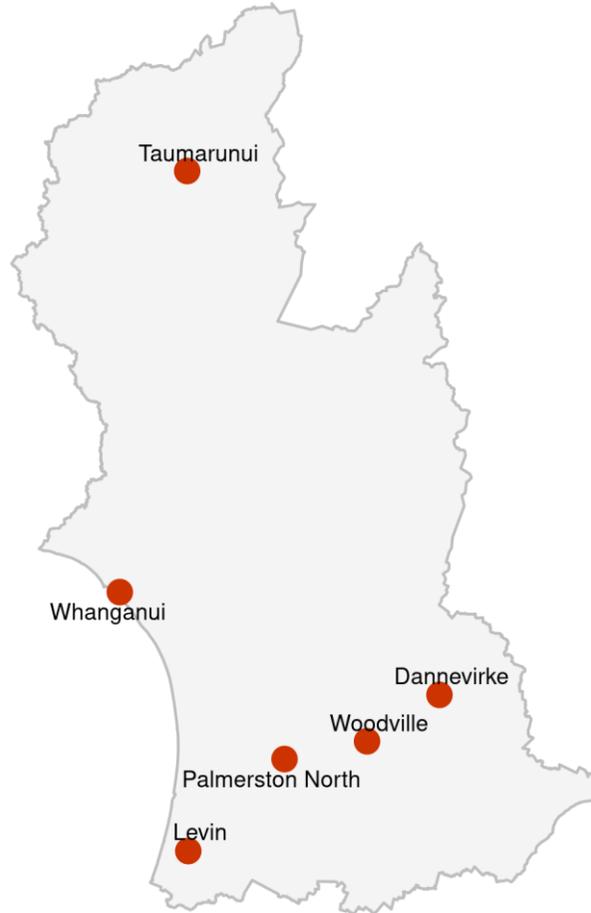
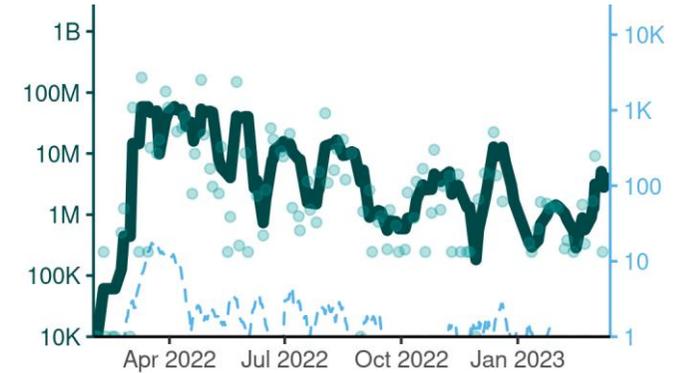
Dannevirke Grab 5.7K



Levin Autosampler 21.2K



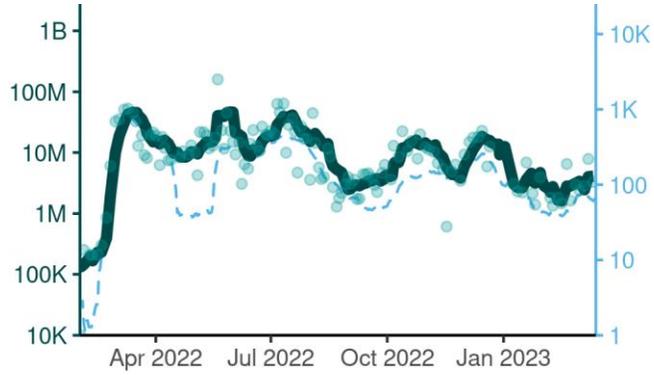
Taumarunui Grab 4.0K



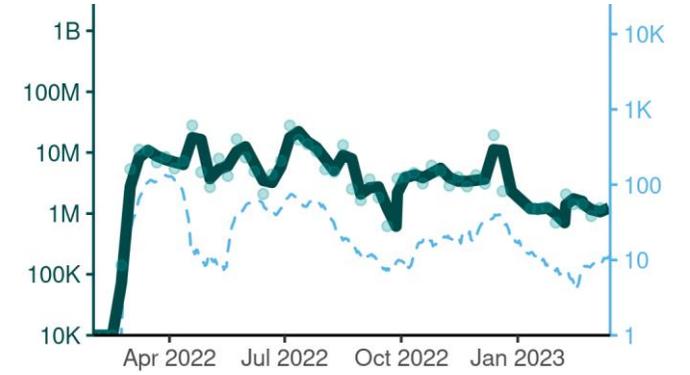
Status ● Detected ● Not detected

Wellington

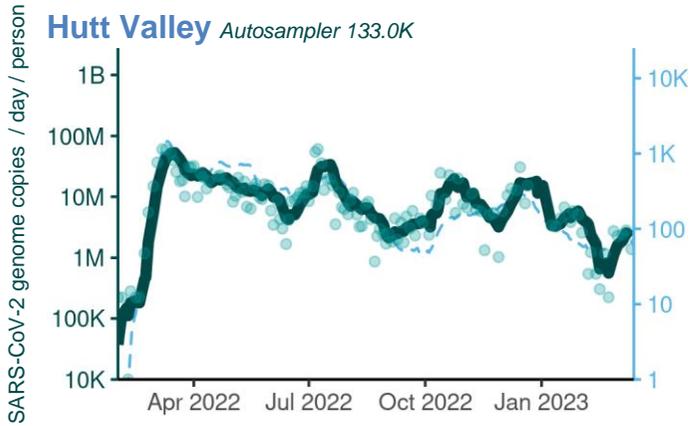
Wellington (Moa Point) Autosampler 168.0K



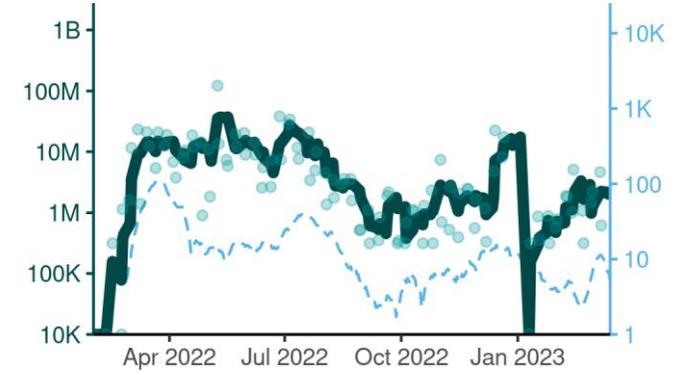
Paraparaumu Autosampler 49.0K



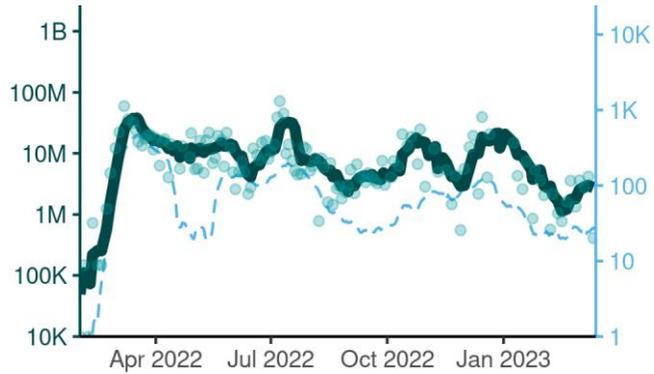
Hutt Valley Autosampler 133.0K



Masterton Auto/grab 20.7K

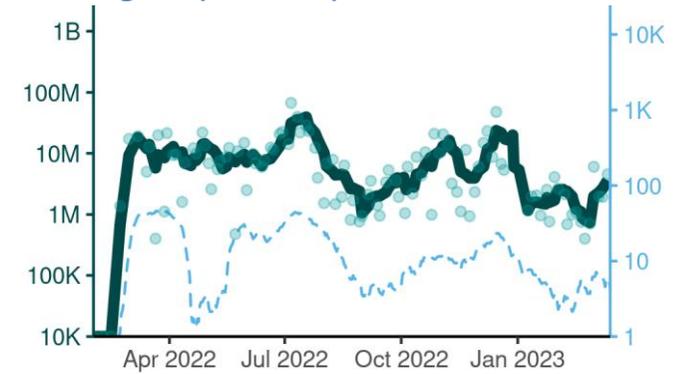


Porirua Autosampler 85.0K



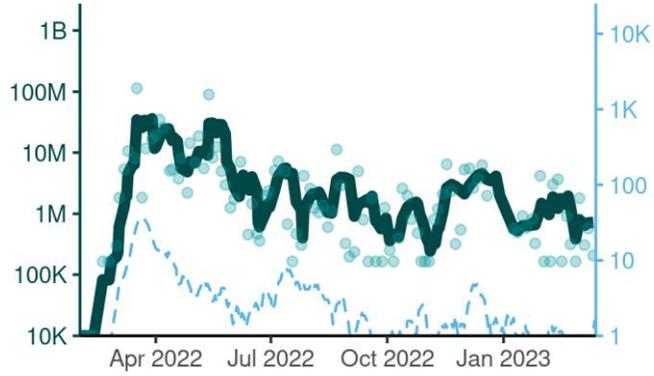
Status ● Detected ● Not detected

Wellington (Western) Autosampler 14.0K

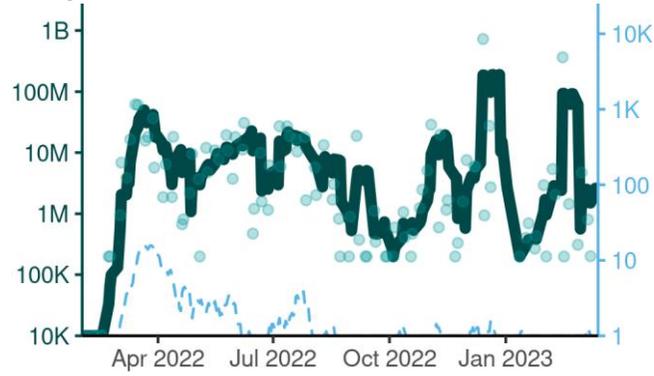


Cases - 7 day rolling average

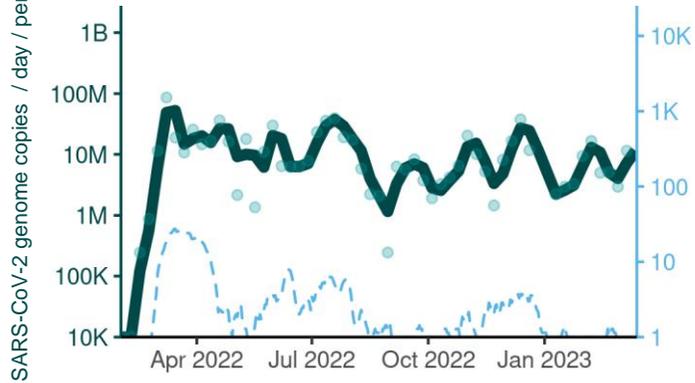
Carterton *Grab 5.8K*



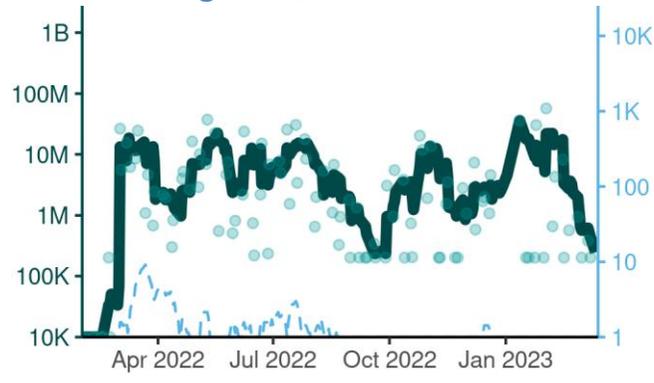
Greytown *Grab 2.4K*



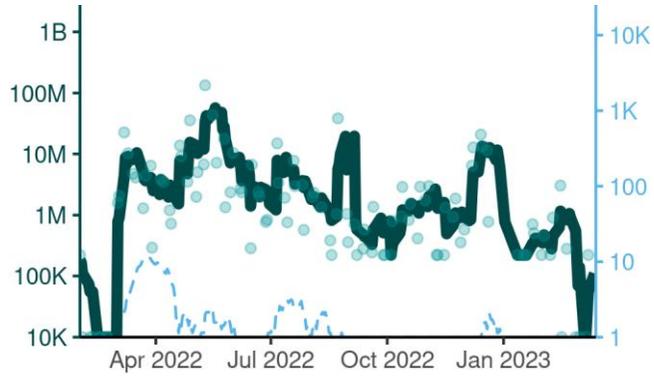
Otaki *Autosampler 3.5K*



Martinborough *Auto/grab 1.6K*

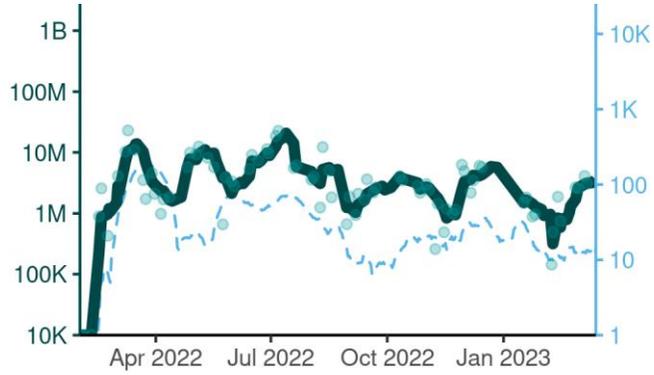


Featherston *Grab 2.5K*

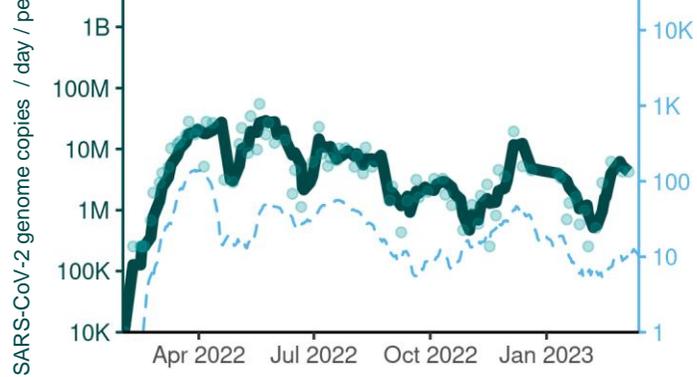


Cases - 7 day rolling average

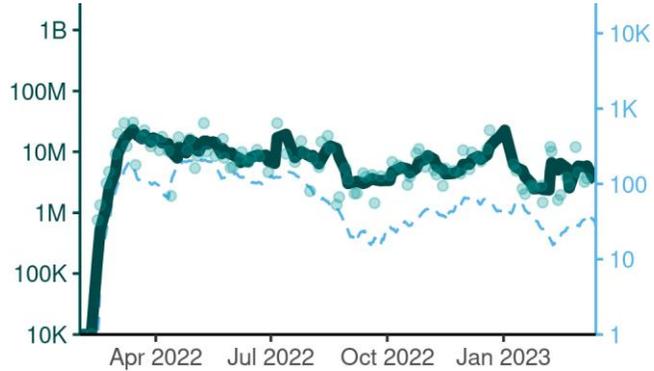
**Tasman, Nelson, and Marlborough
Richmond/Nelson South** Autosampler 60.0K



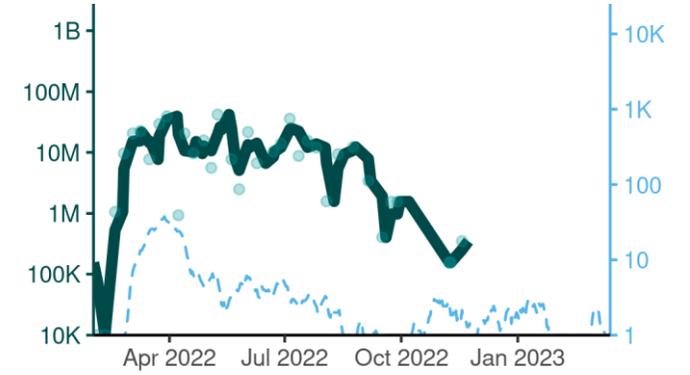
Blenheim Autosampler 31.0K



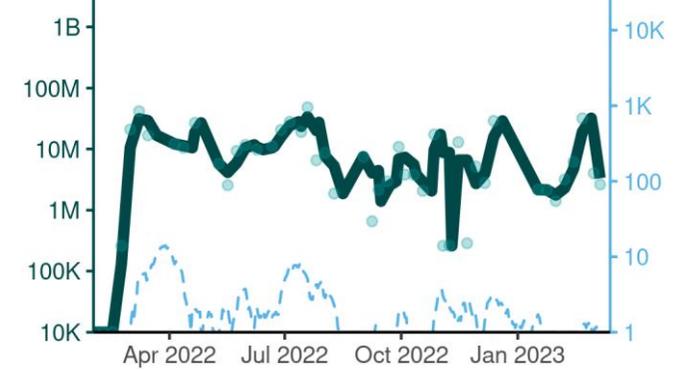
Nelson Central/North Autosampler 26.0K



Motueka Autosampler 8.3K



Picton Autosampler 5.0K

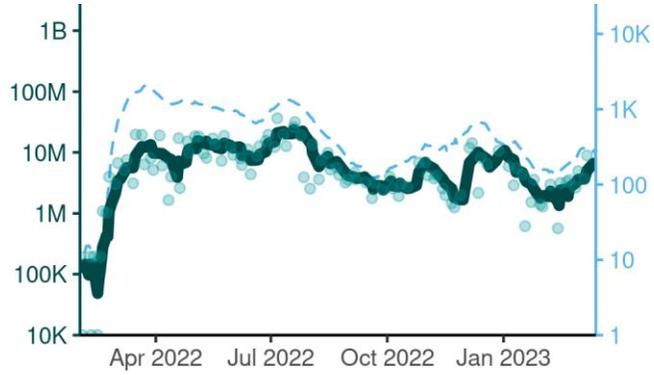


Status ● Detected ● Not detected

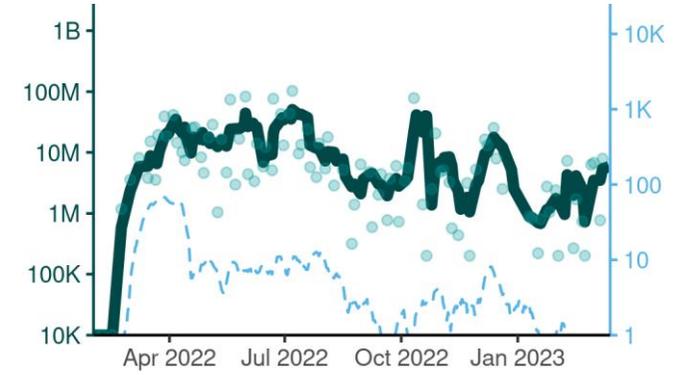
Cases - 7 day rolling average

West Coast and Canterbury

Christchurch Autosampler 368.0K

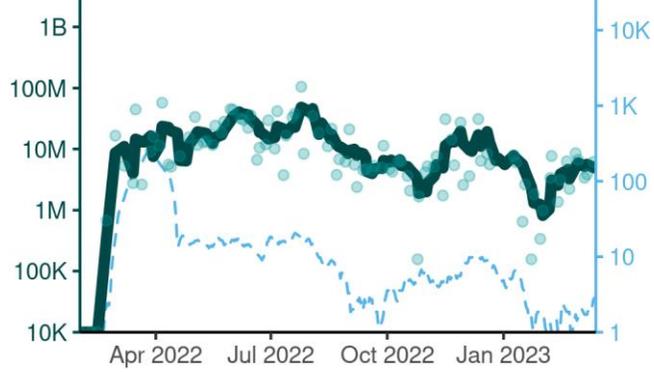


Rangiora Grab 19.0K

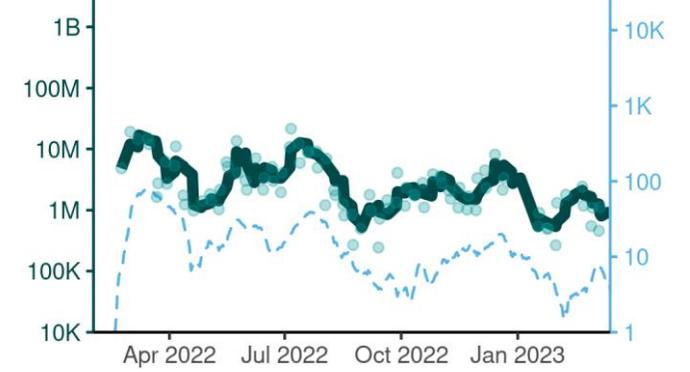


Rolleston & Eastern Selwyn Autosampler 35.0K

SARS-CoV-2 genome copies / day / person

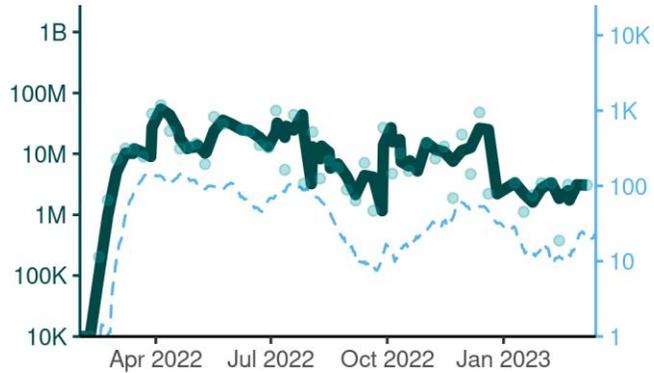


Ashburton Autosampler 18.0K

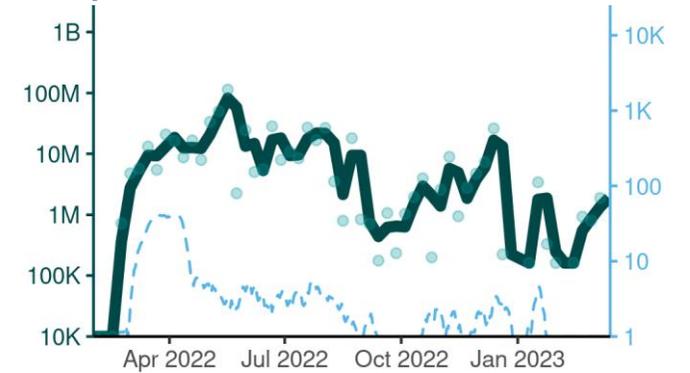


Cases - 7 day rolling average

Timaru Autosampler 28.0K

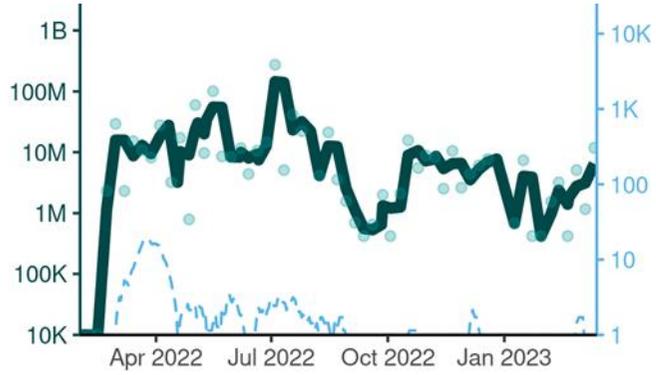


Kaiapoi Grab 12.5K

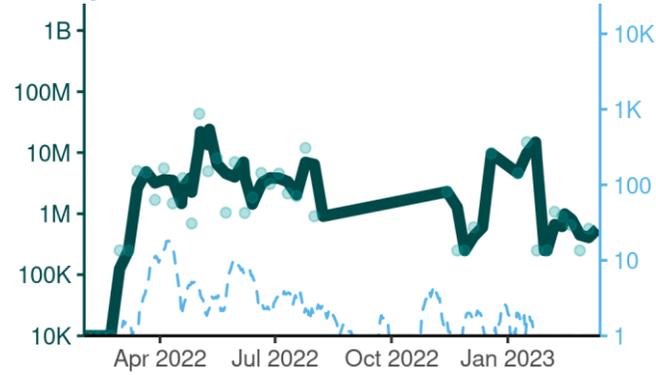


Status ● Detected ● Not detected

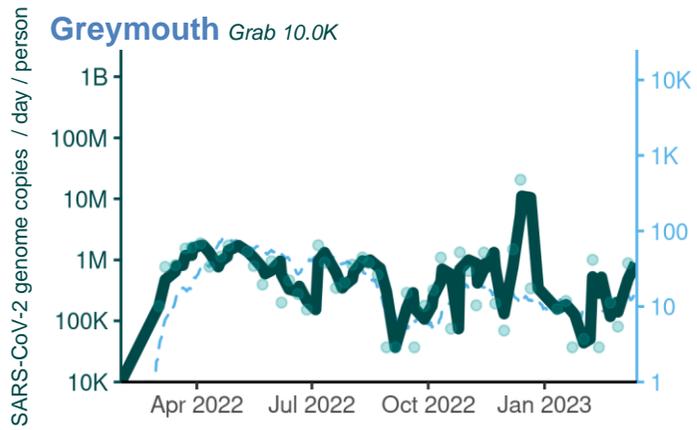
Leeston Autosampler 3.9K



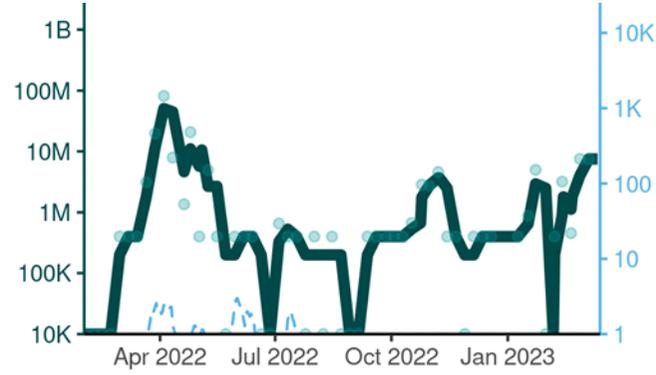
Westport Grab 5.0K



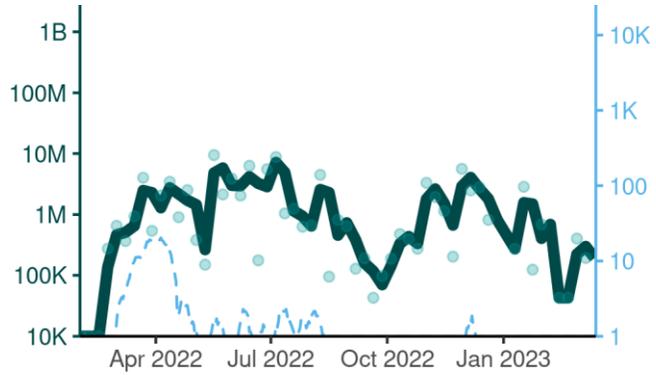
Greymouth Grab 10.0K



Reefton Grab 1000



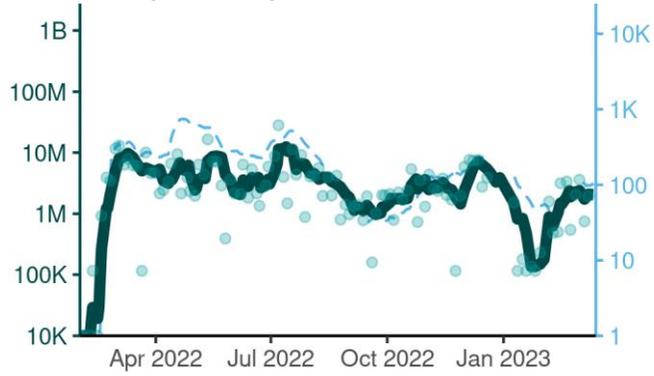
Woodend Grab 7.6K



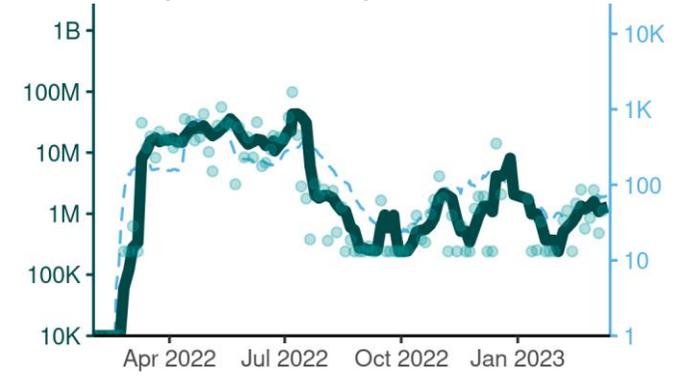
Otago and Southland

Cases - 7 day rolling average

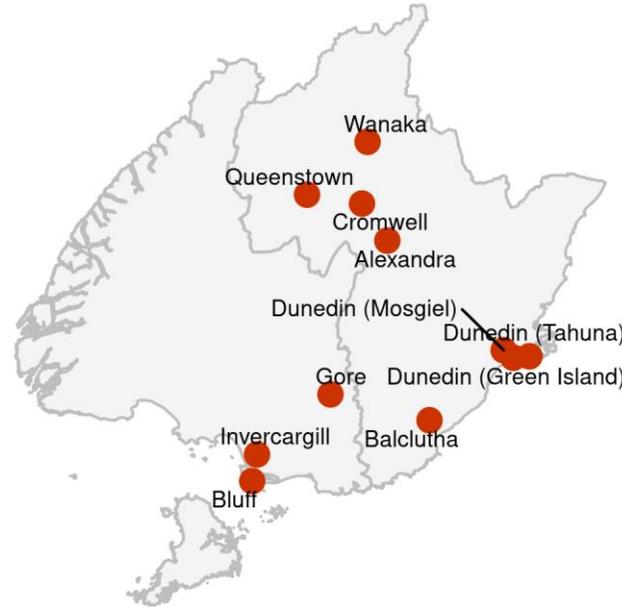
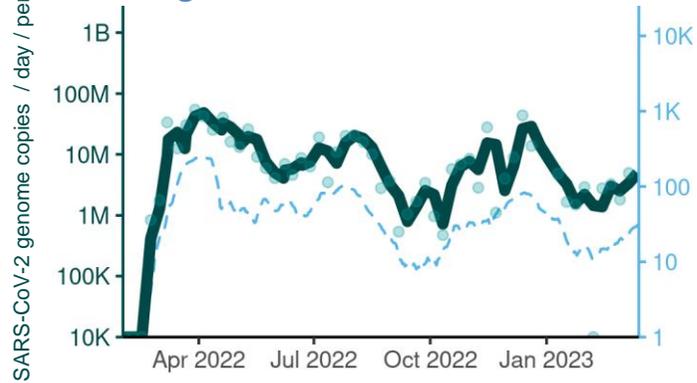
Dunedin (Tahuna) Autosampler 84.0K



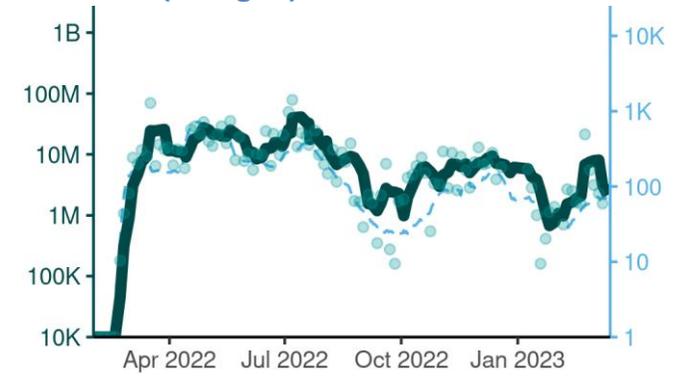
Dunedin (Green Island) Autosampler 22.9K



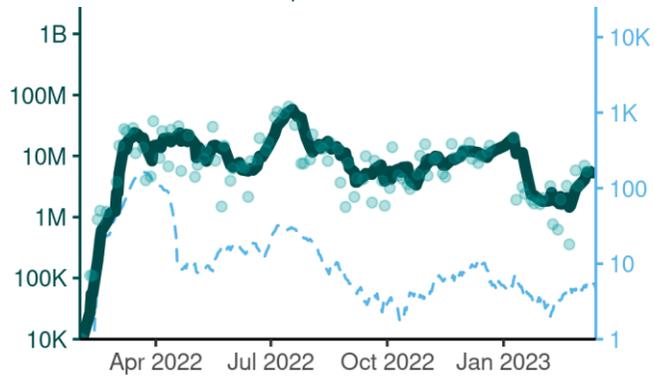
Invercargill Autosampler 50.0K



Dunedin (Mosgiel) Autosampler 14.6K

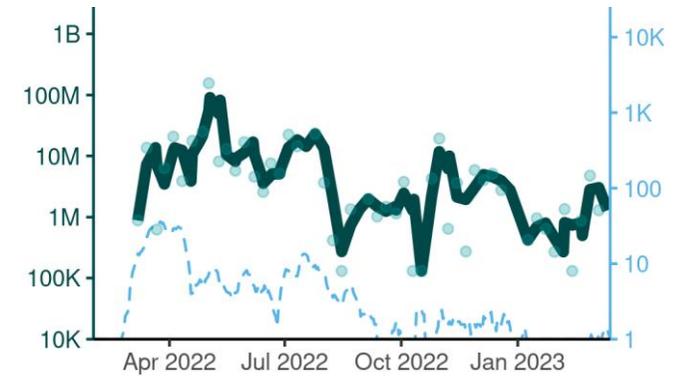


Queenstown Autosampler 40.0K



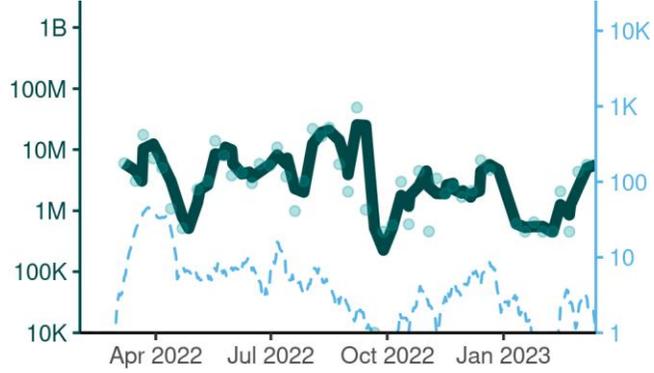
Status ● Detected ● Not detected

Wanaka Grab 14.5K

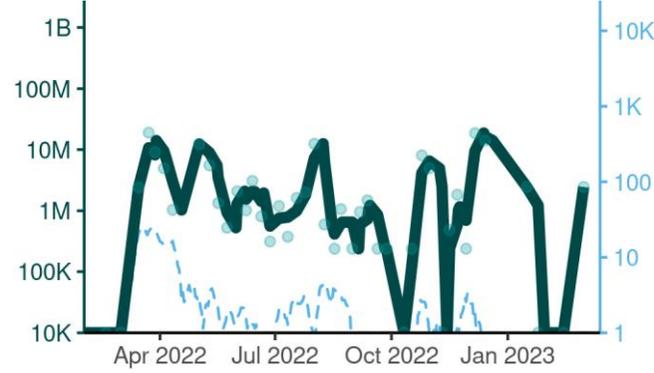


Cases - 7 day rolling average

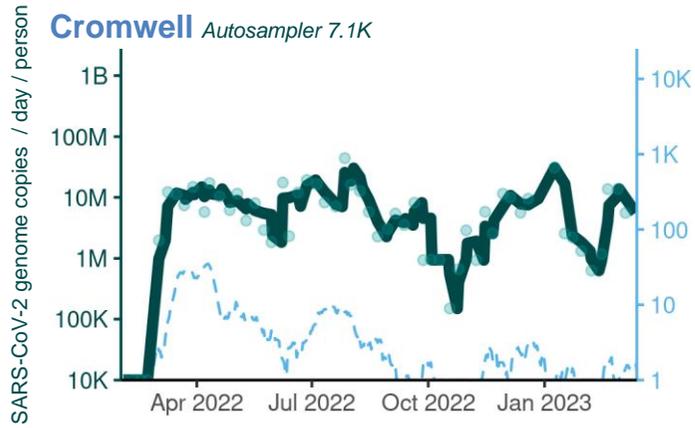
Gore Autosampler 8.0K



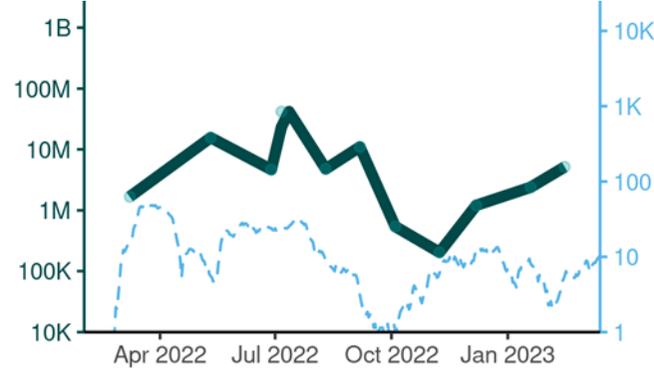
Balclutha Grab 4.1K



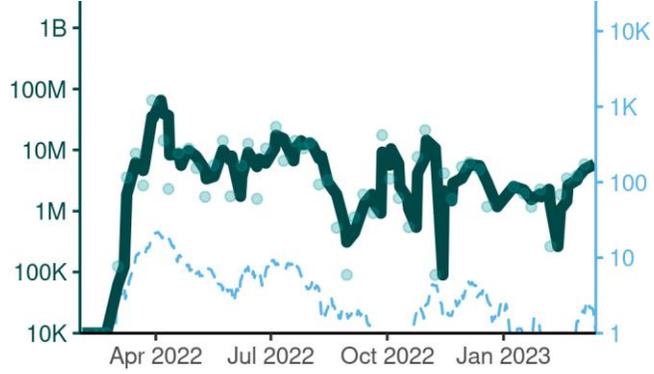
Cromwell Autosampler 7.1K



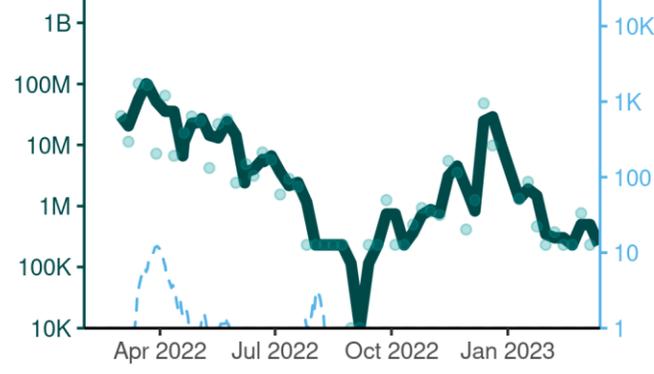
Bluff Autosampler 2.0K



Alexandra Autosampler 6.2K



Oamaru Autosampler 12.0K



Cases - 7 day rolling average

Table 2: Results for weeks 9 & 10 (ending 5 March and 12 March 2023)

Wastewater testing results. Grab samples are collected usually over 15-30 minutes. Autosampler are 24-hour composites.

Region	Site	Population	Sample Type	February 2023					March 2023				
				Mon 27	Tue 28	Wed 01	Thu 02	Fri 03	Mon 06	Tue 07	Wed 08	Thu 09	Fri 10
Northland	Dargaville	5,000	Grab			Detected				Detected			
	Hikurangi	1,730	Grab	Detected						Detected			
	Kaiwaka	400	Grab							Detected			
	Mangawhai	1,100	Grab							Detected			
	Maungaturoto	1,300	Grab							Detected			
	Ruakaka	4,500	Grab	Detected							Detected		
	Whangarei	65,000	Autosampler	Detected							Detected	Detected	
Auckland	Army Bay	42,000	Autosampler		Detected								
	Auckland East	680,000	Autosampler	Detected						Detected		Detected	
	Auckland West	315,000	Autosampler							Detected		Detected	
	Beachlands	6,760	Grab			Detected						Detected	
	Helensville	3,800	Autosampler		Detected						Detected		
	Kawakawa Bay	600	Grab			Detected						Detected	
	North Shore	240,000	Autosampler	Detected		Detected				Detected		Detected	
	Pukekohe	20,900	Autosampler									Detected	
	Snells/Algies	4,000	Autosampler		Detected						Detected		
	Warkworth	3,500	Autosampler		Detected						Detected		
	Waikato	Cambridge	20,100	Autosampler			Detected						
Hamilton		169,000	Autosampler	Detected	Detected					Detected	Detected		
Taupo		23,000	Auto/grab		Detected		Detected				Detected		Detected
Te Awamutu		13,100	Autosampler			Detected							Detected
Thames		7,500	Autosampler										Detected
Whangamata		4,000	Autosampler								Detected		
Whitianga		6,600	Autosampler								Detected		
Bay of Plenty	Katikati	5,500	Autosampler			Detected						Not detected	
	Kawerau	7,000	Autosampler		Detected							Detected	
	Maketu	1,300	Autosampler									Detected	
	Mt Maunganui/Papamoa	65,000	Autosampler		Detected		Detected				Detected		Detected
	Opotiki	3,800	Autosampler	Detected			Detected				Detected		Detected
	Rotorua	59,000	Autosampler		Detected		Detected				Detected		Detected
	Tauranga	50,000	Autosampler		Detected		Detected				Detected		Detected
	Te Puke	9,700	Autosampler									Detected	
	Waihi Beach	3,600	Autosampler			Detected						Detected	
	Whakatane	21,020	Autosampler		Detected						Detected		
Hawke's Bay	Hastings	80,000	Autosampler			Detected	Detected			Detected	Detected		
	Waipukurau	4,610	Autosampler				Detected						Detected
Taranaki	Eltham	2,006	Autosampler			Detected						Detected	
	Hawera	12,000	Autosampler		Detected	Detected					Detected	Detected	
	New Plymouth	88,000	Autosampler				Detected				Detected		Detected
Manawatu-Whanganui	Dannevirke	5,696	Grab				Detected						Detected
	Levin	21,200	Autosampler		Detected		Detected				Detected		Detected
	Palmerston North	90,000	Autosampler		Detected		Detected				Detected		Detected
	Taumarunui	4,000	Grab			Detected		Detected					Detected
	Whanganui	44,500	Autosampler		Detected		Detected				Detected		Detected
	Woodville	1,657	Grab				Detected						Detected
Wellington	Carterton	5,800	Grab	Detected			Detected			Detected			Detected

Region	Site	Population	Sample Type	February 2023					March 2023				
				Mon 27	Tue 28	Wed 01	Thu 02	Fri 03	Mon 06	Tue 07	Wed 08	Thu 09	Fri 10
	Featherston	2,500	Grab	Not detected			Not detected			Not detected		Detected	
	Greytown	2,438	Grab	Detected			Detected			Detected		Detected	
	Hutt Valley	133,000	Autosampler	Detected			Detected		Detected			Detected	
	Martinborough	1,641	Auto/grab	Detected			Detected			Detected		Detected	
	Masterton	20,700	Auto/grab		Detected	Detected				Detected	Detected		
	Otaki	3,500	Autosampler		Detected					Detected			
	Paraparaumu	49,000	Autosampler		Detected					Detected			
	Porirua	85,000	Autosampler	Detected			Detected		Detected			Detected	
	Wellington (Moa Point)	168,000	Autosampler	Detected			Detected		Detected			Detected	
	Wellington (Western)	14,000	Autosampler	Detected			Detected		Detected			Detected	
Nelson	Nelson Central/North	26,000	Autosampler	Detected		Detected			Detected		Detected		
	Richmond/Nelson South	60,000	Autosampler	Detected		Detected			Detected		Detected		
Marlborough	Blenheim	31,000	Autosampler				Detected			Detected			
	Picton	5,000	Autosampler				Detected			Detected			
West Coast	Greymouth	10,000	Grab		Detected						Detected		
	Reefton	1,000	Grab	Detected					Detected				
	Westport	5,000	Grab	Detected					Detected				
Canterbury	Ashburton	18,000	Autosampler		Detected	Detected			Detected	Detected			
	Christchurch	368,000	Autosampler	Detected		Detected			Detected		Detected		
	Kaipoi	12,500	Grab		Detected					Detected			
	Leeston	3,900	Autosampler	Detected					Detected				
	Rangiora	19,000	Grab		Detected		Detected			Detected		Detected	
	Rolleston & Eastern Selwyn	35,000	Autosampler	Detected		Detected			Detected		Detected		
	Timaru	28,000	Autosampler								Detected		
	Woodend	7,600	Grab		Detected					Detected			
Otago	Alexandra	6,200	Autosampler		Detected				Detected				
	Balclutha	4,100	Grab				Detected						
	Cromwell	7,100	Autosampler		Detected				Detected				
	Dunedin (Green Island)	22,900	Autosampler	Detected			Detected		Detected			Detected	
	Dunedin (Mosgiel)	14,600	Autosampler	Detected			Detected		Detected			Detected	
	Dunedin (Tahuna)	84,000	Autosampler	Detected			Detected		Detected			Detected	
	Queenstown	40,000	Autosampler	Detected		Detected			Detected		Detected		
	Wanaka	14,500	Grab	Detected					Detected				
Southland	Bluff	2,000	Autosampler		Detected					Detected			
	Gore	8,000	Autosampler			Detected					Detected		
	Invercargill	50,000	Autosampler		Detected					Detected			

Acknowledgements

This work represents the combined efforts of a large number of individuals and organisations.

We continue to be indebted to the teams across the country who are collecting the wastewater that underpins this work.

The wastewater analysis has been undertaken at ESR by a team which may on any given week include contributions from Joanne Chapman, Dawn Croucher, Joanne Hewitt, Joycelyn Ho, Anower Javed, Olivia Macrae, Ashley McDonald, Andrew Ng, Ashley Orton, and Fatiha Sulthana. Data science analysis, visualisation and reporting is the result of team effort from: Franco Andrews, Bridget Armstrong, Raewyn Campbell, Joanne Chapman, Lei Chen, Gerhard de Beer, Richard Dean, Brent Gilpin, Joanne Hewitt, Dawen Li, Jonathan Marshall, Helen Morris and Leighton Watson. Ongoing support for this work from the Ministry of Health and ESR management is appreciated.

Notes

Sites and frequency of sample collection: The catchment population sites selected for the surveillance range from approximately 400 to over 1,000,000 individuals. The sites cover all regions of the country. Most major towns and all cities, as well as many smaller communities, are included. In early 2023, the wastewater catchment areas cover over 75% of the population connected to wastewater treatment plants. The sites from which samples have been collected have varied over the last 12 months. New sites may be added over time, and/or sampling may reduce in frequency or cease for other sites. The selection and frequency of sampling vary depending on the local population, access to wastewater collection points, staff availability to collect samples and risk factors. When included, samples are collected at least weekly, with twice weekly sampling being common.

Sampling method: The preferred option is to automatically collect a 24 hour 'composite' sample. This is where a pump automatically collects a small volume of wastewater every 15 minutes over 24 hours using a composite sampler. These samplers are available in some wastewater treatment plants. When composite samplers are not available, 'grab' samples are collected. These range from a sample being taken at a single point in time, to 3 samples taken over 30 minutes, to samples collected over a day. Grab samples represent only the composition of the source at that time of collection and may not be as representative as a 24-hour composite sampler. More variation may be expected with grab samples.

Laboratory analysis of wastewater samples: Samples are sent from each wastewater treatment plant to ESR. Processing of each sample commences within an hour or two of receipt. Processing involves the concentration of virus from 250 mL sample to approx. 1 mL using centrifugation and polyethylene glycol. Viral RNA is then extracted from a small volume of 0.2 mL concentrate to give a final volume of 0.05 mL. The presence of SARS-CoV-2 RNA is determined using RT-qPCR. SARS-CoV-2 is considered detected when any of the RT-qPCR replicates are positive.

RT-qPCR: Reverse transcription (RT) to convert RNA to complementary DNA (cDNA), followed by quantitative PCR (qPCR). RT-qPCR is used for detection and quantification of viral RNA.

Method sensitivity: The protocol used to concentrate SARS-CoV-2 from wastewater allows for the sensitive detection of SARS-CoV-2 by RT-qPCR. ESR has shown that when 10 individuals are actively shedding SARS-CoV-2 RNA in a catchment of 100,000 individuals, there was a high likelihood of detecting viral RNA in wastewater (<https://doi.org/10.1016/j.watres.2021.118032>). Shedding by one individual may be detected in wastewater, but it does depend on many factors including the amount and duration of shedding. Very low levels in wastewater may be not able to be quantified (i.e., less than the limit of quantification- see below).

SARS-CoV-2 RNA detected (positive result): A positive detection in the wastewater indicates that at least one person has been shedding SARS-CoV-2 into the wastewater at some point during the time period that the sample was being collected. In some cases, detections could also be due to the shedding of low levels of SARS-CoV-2 RNA by a recently recovered case. The detection of SARS-CoV-2 RNA does not indicate that infectious virus is present.

SARS-CoV-2 RNA not detected (negative result): A negative result can occur because there are no active 'shedding' cases in the catchment or because the SARS-CoV-2 RNA concentration is too low to be detected, most likely because there are a very low number of cases in the wastewater catchment. Therefore, negative finding does not necessarily guarantee the absence of COVID-19 in the community.

Viral loads and normalisation: When detected, the SARS-CoV-2 RNA concentration is calculated as genome copies per L of wastewater. This is then converted to a viral load of **genome copies/day/person**. This conversion takes into account the flow rate of wastewater entering the treatment plant (the influent) and the population in the catchment. The **flow rate** is the total volume (m³ per day) recorded at the inlet of the wastewater treatment plant over 24 hours. This is a **population-normalised viral load**. Currently, the flow rate is the average annual flow rate, but will be replaced with daily flow rate when available (note that rainfall may significantly increase the flow rate at the inlet, diluting the sample, and may result in lower concentrations and a false negative result).

Limit of quantification: The lowest concentration of the target that can be reliably quantified is referred to as the limit of quantification. For those samples where SARS-CoV-2 is detected but cannot be quantified, a value of 5 genome copies/mL wastewater is used. While a standard method is being used, virus recovery can vary from sample to sample, and this may affect the quantitation.

Wastewater Data Modelling

COVID-19 surveillance in Aotearoa, based on self-reported tests, can be problematic due to decreased testing and reporting. This difficulty is further amplified by increasing numbers of asymptomatic infections and reinfections. Accordingly, there is a need to use additional tools to monitor the rates/trends of infection within the community and to explore what information, predictive potential, and uncertainty, lies within the wastewater data. To address this, ESR has partnered with Auckland, Massey and Canterbury Universities (including the COVID-19 Modelling Aotearoa group; CMA) to address these issues. Modelling infection dynamics using wastewater data has challenges. There are many confounding factors that can influence the levels of SARS-CoV-2 in a wastewater catchment - from rainfall effects and shedding rates through to differences in RNA concentrations between variants. Current modelling reports on two key metrics (i) an estimate of the instantaneous reproduction number 'R' based on both case data and wastewater quantitation and (ii) an estimate of the case ascertainment rate (CAR).

Data subject to change: Data generated for the New Zealand Wastewater COVID-19 Surveillance Programme should be considered provisional and may be subject to change. Data may be incomplete for the most recent 2-week period due to processing, testing and reporting delays.

Data not shown:

- Results from certain samples may not be shown, as the result was either deemed invalid, or the sample could not be tested (e.g., leaked in transit, not labelled).

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